



Thermoregulation in the wild boar (*Sus scrofa*)

Thomas Ruf¹ · Sebastian G. Vetter^{1,2} · Johanna Painer-Gigler¹ · Gabrielle Stalder¹ · Claudia Bieber¹

Received: 2 June 2023 / Revised: 25 August 2023 / Accepted: 5 September 2023 / Published online: 24 September 2023
© The Author(s) 2023

Abstract

The wild boar (*Sus scrofa*) originates from warm islands but now inhabits large areas of the world, with Antarctica as the only continent not inhabited by this species. One might be tempted to think that its wide distribution results from increasing environmental temperatures. However, any effect of temperature is only indirect: Abundant availability of critical food resources can fully compensate the negative effects of cold winters on population growth. Here, we asked if temperature as a habitat factor is unimportant compared with other habitat indices, simply because wild boars are excellent thermoregulators. We found that the thermoneutral zone in summer was approximately 6–24 °C. In winter, the thermoneutral zone was lowered to 0–7 °C. The estimated increase in the heart rate and energy expenditure in the cold was less than 30% per 10 °C temperature decline. This relatively small increase of energy expenditure during cold exposure places the wild boar in the realm of arctic animals, such as the polar bear, whereas tropical mammals raise their energy expenditure several fold. The response of wild boars to high T_a was weak across all seasons. In the heat, wild boars avoid close contact to conspecifics and particularly use wallowing in mud or other wet substrates to cool and prevent hyperthermia. Wild boars also rely on daily cycles, especially of rhythms in subcutaneous temperature that enables them to cheaply build large core–shell gradients, which serve to lower heat loss. We argue it is predominantly this ability which allowed wild boars to inhabit most climatically diverse areas in the world.

Keywords Thermoregulation · TNZ · Heart rate · Core–shell · Subcutaneous temperature · Mass

Introduction

The current distribution of wild boars (*Sus scrofa*) extends to most areas of the world with Antarctica being the only continent not inhabited by this species (Lewis et al. 2017). In fact, no other mammal, except for humans and their companions the house mouse (*Mus musculus*) and the Norwegian rat (*Rattus norvegicus*), has settled in as many climatically diverse areas. Its evolutionary lineage originates from islands in Southeast Asia (Philippines, Indonesia) (Chen

et al. 2007), and in warm countries such as Vietnam, wild boars still thrive and pig farming is an important meat source (Thom et al. 2021). *S. scrofa* naturally expanded throughout Eurasia and into North Africa (Wehr 2021). Remarkably, now the wild boar in Europe is distributed up to 64°N. In Asia, the northern distributional limit is up to 61°N and it was reported to inhabit even the coldest climates in Siberia (Markov et al. 2019).

In fact, the wild boar is not only a widespread species but has become a pest in large areas. In many parts of Europe, densities of wild boars have been growing notably during the past few decades (e.g., Vetter et al. 2015). This increase has led to substantial damage to agricultural crops and can even cause problems in urban habitats (Geisser and Reyer 2005). Feral pigs in Australia, New Zealand, and other southern countries cause similar damage (Dexter 2003; Hampton et al. 2004). The increase in wild boar populations may be the result of the global increase in mean environmental temperatures. However, any effect of ambient temperature (T_a) is only indirect, via food. The abundant availability of critical food resources, e.g., beech nuts, can outweigh the negative

Communicated by G. Heldmaier.

✉ Thomas Ruf
thomas.ruf@vetmeduni.ac.at

¹ Department of Interdisciplinary Life Sciences, Research Institute of Wildlife Ecology, University of Veterinary Medicine, Savoyenstrasse 1, 1160 Vienna, Austria

² Present Address: Department for Farm Animals and Veterinary Public Health, Institute of Animal Welfare Science, University of Veterinary Medicine, Veterinärplatz 1, 1210 Vienna, Austria

effects of cold winters on population growth of wild boar (Vetter et al. 2015).

More recently, the regions of successful settlement of wild boars were analyzed in detail based on 129 records of population densities and habitat characteristics (Lewis et al. 2017). Interestingly, this study showed a remarkable influence of both abiotic and biotic factors, such as large carnivore richness. Ambient temperature seemed to be less important but may play a role on a broad scale (Lewis et al. 2017). However, one may argue that for a species that is excellent at thermoregulation, T_a as a habitat factor may seem unimportant compared with other habitat indices, especially if T_a does not constrain population size. Yet, we are unaware if wild boars are excellent or poor thermoregulators.

Measurements of energy expenditure for thermoregulation were mostly carried out on domestic pigs that are characterized by a thin layer of sparse hairs, and this work often focused on newborn piglets that are especially temperature sensitive (reviewed in Gómez-Prado et al. 2022). In sows, the thermoneutral zone (TNZ) has been reported to be 15–20 °C (Yousef 1985). For an overview on heat stress in pigs, see Brown-Brandl et al. (2004). Measurements on actual wild boar also seem restricted to observations mostly on juveniles (Jeziarski and Myrcha 1975). Part of the reason for the scarcity of metabolic data is that adult wild boars tend to destroy respirometric chambers.

We asked if wild boars are relatively cold tolerant and have a lower TNZ than domesticated pigs. In addition, we were interested in how much wild boars raise their energy expenditure for thermoregulation. Indirectly, we wanted to see if their ability to inhabit most climatically diverse areas of the world is largely due to excellent thermoregulation. We addressed these questions using heart rate measurements. We further implanted temperature loggers at different parts of the body to measure a possible temperature gradient in the body in detail.

Material and methods

Animals and study area

The study animals were kept in an outdoor enclosure (~55 ha) in the Burgenland (Austria). The study enclosure was covered with a deciduous forest, mainly Turkey oak (*Quercus cerris*) and pubescent oak (*Quercus pubescens*) and included a few meadow patches. For the present study, 13 adult females were used. Likely, all females gave birth and were lactating. We concentrated on females only because the live capture and handling of males are hampered by their large size and ferocity. Also, due to competition and high levels of aggression between males during rut, the stocking of the enclosure was strongly female biased. During

the study period (12/2016–01/2019), the animal density was ~1 adult female/ha plus up to 20 males (total) of different ages. Due to this relatively high density, animals were supplemented with 1–1.5 kg corn/individual once a day (at 2:00–14:00 h) at two feeding areas, each ~40 × 20 m. The enclosure was enclosed by 2.5 m high, solid, non-transparent fencing and was closed for the public. Thus, the study site provided an environment without disturbances due to hikers, bikers or straying dogs. There were no battue hunts or other disturbances due to hunting or forest management activities during the study period in the enclosure.

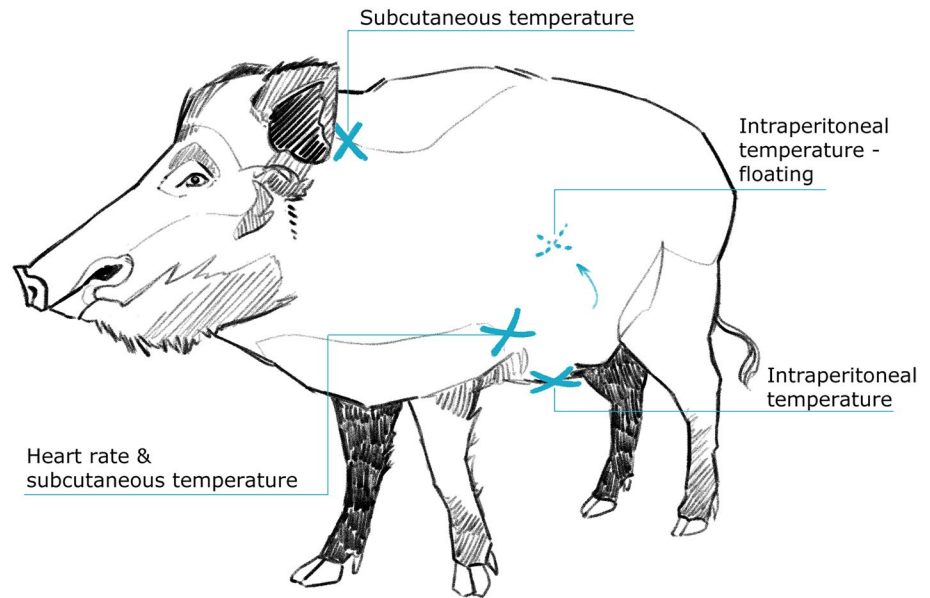
Animals were trapped once a year in autumn within the feeding sites to collect data on reproductive success (collecting DNA of juveniles) and body condition of females and to separate some of them for implantation/explantation of loggers. While feeding, we closed the access gates and released the boar one by one through a wooden corridor back into the enclosure. While in the wooden corridor, we recorded the body mass of each individual (Gallagher SmartScale® 500, Groningen, Netherlands). Due to management reasons, the juveniles (born in spring) were removed from the enclosure during this procedure.

Implantation of temperature and heart rate loggers

We implanted a heart rate and temperature logger (HR&T; DST centi-HRT, Star-Oddi, Gardabaer, Iceland, and custom built at the Research Institute of Wildlife Ecology, Vienna, Austria, Fig. 1). In ten females, we implanted heart rate and temperature loggers at the sternum as well as temperature loggers at the neck. Nine of these females also received intraperitoneal wall temperature loggers, and in the tenth animal, the logger was internally floating (ruptured from the intraperitoneal wall). Three additional females were implanted with intraperitoneal-floating and wall temperature loggers. Two of these three also had subcutaneous neck loggers. All 13 animals underwent surgery in October/November 2016 and 2017 as adults (females aged 5 and 6 years). Logger records lasted up to a little more than one year. All details about surgery techniques and anesthesia protocols were provided before (Ruf et al. 2021). Explantations were carried out approximately one year after implantations. The last explantation was carried out in January 2019. Mean body mass at date of implantation for all females was 71.8 ± 15.5 kg. Body mass was not determined during HR measurements and too rarely to establish an allometry.

The HR&T logger (Fig. 1) sampled data at 12 min intervals without specific limits to maximum and minimum. We did not further validate records. It was positioned subcutaneously, in proximity to the heart on the lateral rib cage, behind the moving area of the elbow, to avoid rubbing, or inserted and tethered into the ventral subperitoneal space caudal of the xiphoid process of the sternum.

Fig. 1 Location of the implantation of loggers on wild boar. Each female investigated had a least two and up to 5 loggers (heart and temperature) implanted



Self-built temperature loggers were covered with inert surgical wax and had a weight of ~8 g. Time interval of recording was also adjusted to 12 min, the accuracy 0.01 °C. One of the temperature loggers had an especially flat shape ($3.4 \times 1.9 \times 0.5$ cm) to fit smoothly into the subcutaneous neck region ($n = 10$). The second temperature logger (diameter = 2.1 cm and height = 1.2 cm) was placed into the intraperitoneal cavity, either tethered at the linea alba ($n = 12$), or freely floating in the cavity ($n = 4$), usually found between two lobes of the liver.

Activity data

To record the activity (ear-tag movements) of animals, a telemetry system (Smartbow System, Zoetis, New Jersey, USA) was installed around the two neighboring feeding areas and two close water ponds in the enclosure. The system consisted of a central solar power and computing station and ten receivers located at the height of 2–3 m. Part of the system were ear-tags (34 g; $52 \times 36 \times 17$ mm, $n = 13$). The accelerometer (located inside ear-tags) measured triaxial acceleration (x , y , z). As an estimate of movements, we computed the total acceleration vector from $\sqrt{x^2 + y^2 + z^2}$.

Climate

We recorded ambient temperature (T_a) and black-bulb temperature (T_{ab}) at 2 m height directly at the study site (Vantage Pro 2 with black-bulb extension, Davis Instruments, Hayward, USA).

Data analysis

We did not assess the influence of environmental conditions in different years, due to logger failures and thus scarcity of heart rates, and all data were pooled for different years (with similar conditions and food available year-round). Together, we analyzed a little more than 0.5 million data sets (with equal contributions from times, heart rates, activities and temperatures).

To remove outliers, all initial data from recorders were subjected to the R package boxfilter that removes isolated data points (Ruf 2022). To investigate the influence of T_a , heart rates were plotted with ggplot2 (Wickham 2016) as a function of T_a for meteorological summer (01 June–31 October) and winter (01 December–31 May). During warm and cold exposure in both seasons, linear regressions were computed using several minimum heart rates per animal and degree centigrade (Fig. 2). To restrict the data analysis to thermoregulation, we deleted 5 out of 495 regression points, with relatively high heart rates (> 50 bpm) when animals were apparently not at rest. The relationship between heart rate at rest and T_a was investigated for summer and winter separately by using the R package segmented (Mugge 2003). This library determines best breakpoints in linear relations, in this case the critical temperatures below and above the TNZ. Suggested breakpoints were rounded to integers (in °C).

The relation between peripheral temperatures and T_a was investigated with general additive models (R package mgcv (Wood 2017), Figs. 2, 3). This function fits non-linear splines to the data, which are penalized for their “wiggleness”, i.e., the number of turning points in the fit. These models were all corrected for the time of day (using circular

Fig. 2 Heart rate as a function of T_a in wild boar during winter (a December–May) and summer (b June–November). Dots are original heart rates. The thermoneutral zone in summer was approximately 6–24 °C (vertical lines). In winter, the thermoneutral zone was approximately 0–7 °C (vertical solid gray lines). Red regression lines show how heart rate increases below thermoneutrality in summer, blue lines depict winter. Linear regressions were computed using several minimum heart rates per animal and degree centigrade

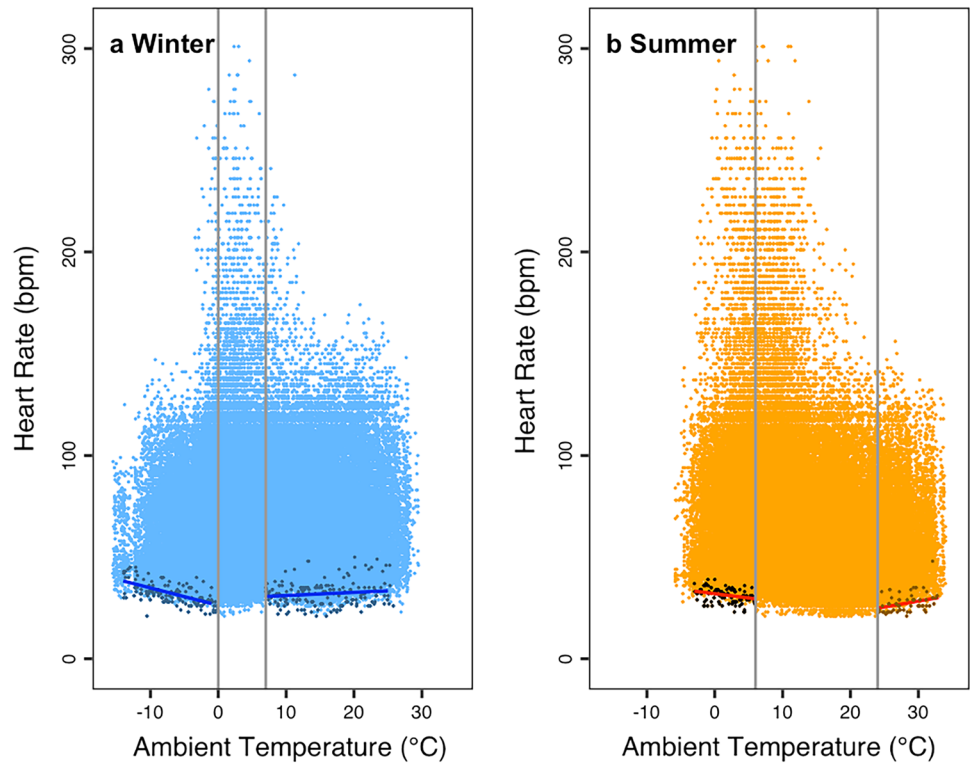
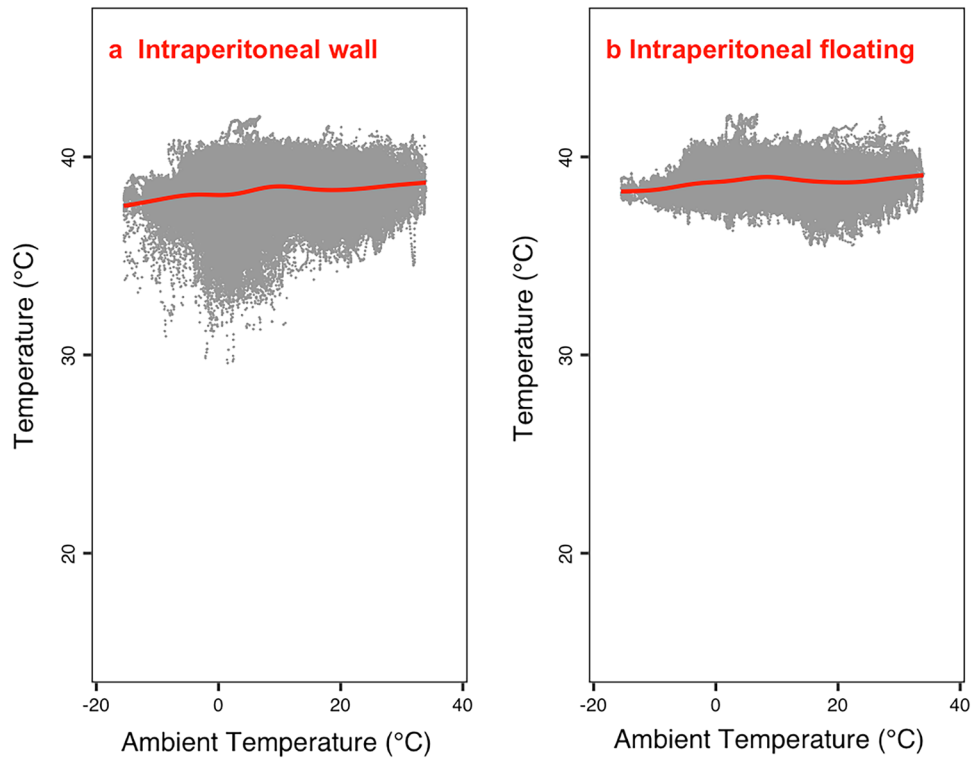


Fig. 3 Intraperitoneal temperature as a function of T_a in wild boar. Temperature loggers measured T_b at 12 min intervals and were either sutured to the intraperitoneal wall (1a) or free floating (found next to the liver, B). GAM models (red lines) indicated that intraperitoneal temperatures decreased with T_a (floating: edf (wiggleness)=8.91, residual df=8.99, $F=867.9$, $P<0.0001$ ***). Partial residual plots corrected for the influence of random differences between individuals. For loggers fixed to the wall, model parameters were (edf (wiggleness)=8.92, residual df=8.99, $F=1030$, $P<0.0001$ ***). All models were also corrected for initial autocorrelation (>0.9)



cubic splines), the random effects of individuals, and for autocorrelation (using rho and the AR.start function).

To illustrate the effects of time of day, means \pm SEM of several measured variables were shown as a function of mean hour (computed using R package lubridate (Grolemund and Wickham 2011)).

Results

The mean heart rate over the entire range of T_a was 66.02 ± 0.051 bpm, approximately three times the minimum heart rate per female (mean: 22.3 ± 0.53). Heart rates were highest (> 200 bpm) at T_a around -2 – 12 °C (Fig. 2). Ambient temperature ranged from -15.5 °C in winter to 34 °C in summer (Fig. 2). There was no appreciable correlation between ear-tag movements and heart rate ($r=0.028$), not even when activity was shifted backward several time intervals (assuming a delayed response of heart rate). Thus, ear-tag movements apparently were not associated with elevated locomotion and heart rate.

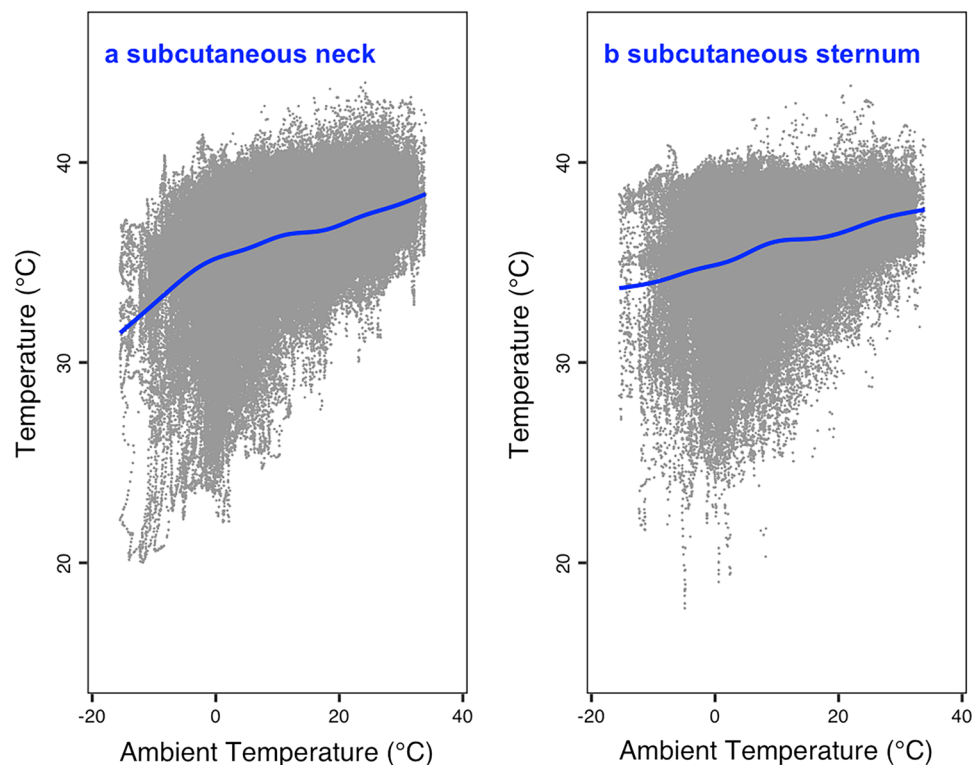
In summer, minimum heart rate increased above 24 °C ($Y=14.87+0.44 \times T_a$) and below 6 °C ($Y=31.85-0.48 \times T_a$). This was calculated from a regression through minimum points per T_a including every animal; Fig. 2).

Accordingly, the apparent thermoneutral zone in summer was approximately 6 – 24 °C (vertical lines). In winter, heart rate increased above 7 °C ($Y=29.86+0.12 \times T_a$) and also below 0 °C ($Y=26.28-0.85 \times T_a$) (Fig. 2). Thus, in winter, the thermoneutral zone was 0 – 7 °C. Wild boar increased their minimum heart rate by 13.2% in summer and 28.4% in winter per additional 10 °C at low T_a s. At temperatures above the apparent TNZ, the increase was 25.9% in summer and merely 4.0% in winter.

As T_a decreased, floating-intraperitoneal-temperature stayed fairly stable around 38 °C (Fig. 3). In loggers fixed to the peritoneal wall, however, temperature dropped to a minimum of ~ 32 °C around a T_a of ~ 0 °C. Below 0 °C, temperature at the peritoneal wall increased again. Subcutaneous temperature both at the neck and the sternum decreased to ~ 25 °C as T_a dropped to -15 °C (Fig. 4). The GAM models showed an almost identical decline of both temperatures (Fig. 4). Again, minimum subcutaneous temperature increased as T_a fell below ~ 0 °C (Fig. 4). Overall, especially at low T_a , there was a temperature gradient with intraperitoneal-floating temperature greater than wall temperature, which was in turn greater than subcutaneous temperature.

Mean subcutaneous temperature also showed a daily rhythm that peaked together with mean heart rate in the afternoon (Fig. 5).

Fig. 4 Subcutaneous temperature as a function of T_a in wild boar. Temperature loggers measured T_b at 12 min intervals and were either fixed to the lateral neck (a) next to the sternum (b). GAM models (blue lines) indicated that subcutaneous temperatures decreased with T_a (neck: edf (wiggliness)=8.85, residual df=8.99, $F=6281$, $P<0.0001$ ***; sternum: edf (wiggliness)=8.54, residual df=8.94, $F=1285$, $P<0.0001$ ***). Partial residual plots corrected for the effects of random differences between individuals. All models were also corrected for initial autocorrelation (>0.9)



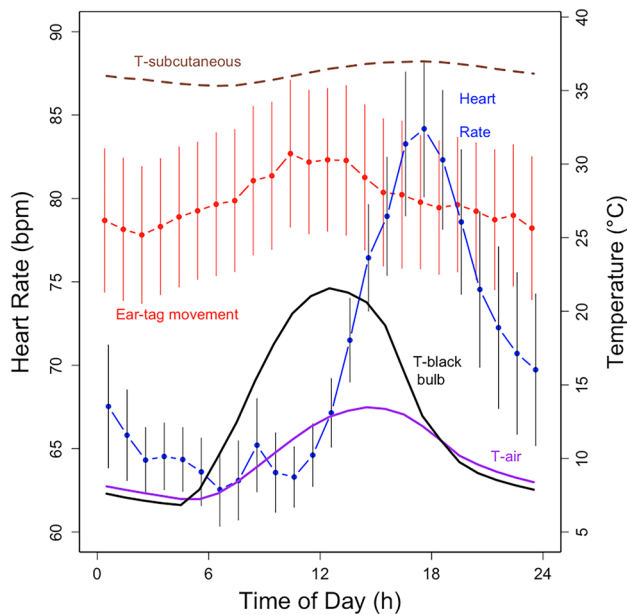


Fig. 5 Hourly means of heart rate (blue), ear-tag movements (red; scale not shown) as well as subcutaneous T_b (brown) as function of daytime in female wild boar under natural conditions in Austria. Further, ambient (purple) and black-bulb temperatures (black) are shown. The SEM on the mean ear-tag movement and the heart rates indicates the variation among 13 animals, the number of time points of each animal varies. This graph is meant to illustrate the daily rhythm of each variable

Discussion

Heart rate as a proxy

Since each heart beat transports oxygen to tissues, it positively correlates with metabolic rate (MR) under most circumstances (Green 2011). Simultaneous measurements of both variables are rare and usually restricted to small species, but at any physiological state their relation is typically linear (e.g., Butler et al. 2004; Currie et al. 2014; Green 2011). However, here we do not aim to calibrate heart rate, we merely assume that effects of T_a on heart rate equally influence MR.

The average heart rate was only about three times that of minimum heart rate. Hence, the wild boar females in this study, which had ample food in addition to their natural forage, were far from maxima observed in mammals (i.e., limits of existence), which can reach up to 12 times basal metabolic rate (Mellish et al. 2000). We also consider energy costs for minimum heart rate during thermoregulation to be only minor (Fig. 2). The missing correlation between “activity” and heart rate suggests that movements of the ears may be a proxy of general alertness but cause no measurable change in energy expenditure.

The only previous observations on thermoregulation in adult wild boar were measurements of MR at various T_a in different age-groups (Jeziński and Myrcha 1975). The overall picture obtained was very similar to our observations, with increases below 0 °C (in 11 month old boar) and above +6 °C (here: +7 °C), but measurements of adults in or below the TNZ seemed to be missing (Jeziński and Myrcha 1975).

Responses to low T_a

Generally, wild boar benefit from their large body size and their near spherical shape, which minimizes the area of heat loss. Vetter et al. (2015) found that body mass among adults correlates negatively with long-term regional mean winter temperatures across Europe. Wild boar in the coldest region were about 30–40 kg heavier than in the warmest region. These differences in the body size contribute to the static advantages of wild boar in the cold.

The observed shift in the TNZ from summer to winter was likely due to improved thermal conductance. The obvious, but theoretical, part of this change was the development of a denser winter fur. In winter, the increase in the heart rate in the cold below the TNZ was less than 30% per 10 °C T_a decline. This relatively small increase in the heart rate during cold exposure places wild boar in the realm of arctic animals, such as the polar bear, whereas tropical mammals raise their energy expenditure several fold (Heldmaier et al. 2013). We suggest that this resistance is largely due to the development of low peripheral temperatures (Fig. 4), i.e., a pronounced body shell. Low peripheral temperatures decrease the body surface to environment gradient and thus limit heat loss. Temperature gradients in the body belong to the most important adaptations to life in the cold (Blix 2016). The ability to control surface temperature and to regulate heat loss was previously shown to increase with body size (Phillips and Heath 1995). Thus, it is not surprising that it occurs in the wild boar. The fact that these gradients are reduced again at even lower T_a , maybe due to the avoidance of adverse effects of low T_a on the skin, underlines that these gradients result from active regulation rather than an unavoidable cooling of peripheral tissues.

In this context, it should be noted that clades ancestral to the *Suidae* lost the heat generating protein UCP1 in non-shivering thermogenesis (NST) about 40 million years ago (Fyda et al. 2020), as it was lost in many mammalian lineages. UCP1 has been mainly abandoned in mammals of large body size, strong increases in body mass followed each loss of UCP1, and NST capacity is inversely related to body size (Gaudry et al. 2017). Although functional UCP1 was retained by some large mammals (Gaudry et al. 2017), we suggest that a decrease in the role of NST in large animals was partly due to their ability to form large core-shell

gradients. Indeed, this is the first time that we can provide a functional explanation for the link between UCP1 loss and large body mass.

One might be tempted to think that in wild boars NST, in the absence of UCP1, plays no role. However, another type of NST, driven by the SERCA pump in muscles, enables juvenile wild boars to generate heat, while shivering is diminishing (Nowack et al. 2019). This type of NST is also present in pigs, the domestic form of wild boar, and seems to be widespread even within adult mammals (Berthon et al. 1994; Nowack et al. 2017). Thus, NST may have contributed to the T_a induced thermogenesis observed here.

Responses to high T_a s

It seems that energy expenses for heat-avoiding thermoregulatory behavior, that actually cut into energy expenses for foraging, occur in wild boar under heat stress that is more severe than what our animals experienced. The observed response of heart rate to high T_a was weak in wild boar across all seasons (Fig. 2). It seems that this very moderate rise in the heart rate was largely due to the choice of a forest as the preferred habitat. This choice of shady areas avoids extreme temperatures that occur in open habitats (for example, see black-bulb temperature in Fig. 5). Wild boar largely lack functional sweat glands for evaporative cooling (Renaudeau et al. 2006) and were seen panting in the field only under severe heat stress (C.B. pers. observation). Only when the hypothalamic temperature is elevated to ~ 39 °C do pigs engage in panting (Baldwin and Ingram 1968). Generally, wild boar and pigs react to changes in temperature, radiation, humidity or wind by adjusting their behavior (Bracke 2011; Olczak et al. 2015). In the heat, pigs and wild boar avoid close contact to conspecifics and particularly use wallowing in mud or other wet substrates to cool and prevent hyperthermia (Bracke 2011; Olczak et al. 2015). Also, it has been argued that wild boars are mainly nocturnal (but see Fig. 5) to lower costs for thermoregulation (e.g., Brivio et al. 2017; Johann et al. 2020).

Low costs for thermoregulation in the heat are also supported by the passive daily rewarming of subcutaneous temperature. Wild boar make extensive use of basking in the sun (Ruf et al. 2021). On days with above average solar radiation, they passively warm peripheral tissues diurnally and thereby save 10% of total energy (Ruf et al. 2021). Arguably, it is thus their diurnality, which allows wild boar to establish low body-to-environment gradients cheaply. Although increased activity and high heart rates during the day (Fig. 5) were probably facilitated by our feeding in the afternoon. We believe observations of nocturnal activity in wild boar are mainly due to their avoidance of human activities, namely hunting (e.g., Brivio et al. 2017; Johann et al. 2020).

General conclusions

These findings point to the importance of daily rhythms (Fig. 5). It is the reliance on the repeatability of daily cycles, especially of rhythms in subcutaneous temperature that enable wild boar to build large core–shell gradients, which serve to lower heat loss in the cold. Taken together, it seems that the resulting low costs during thermoregulation in the cold act together with effective measures to counteract heat by behavioral strategies in the warm. We argue it is predominantly this ability which allowed wild boar to inhabit most climatically diverse areas (Lewis et al. 2017). We would not be surprised if wild boar showed only small responses to increasing global climate change. However, wild boar are omnivorous, and increasing drought might lead to lower food availability, since digging up roots and insects from below ground seems hardly feasible (Ruf et al. 2021).

Acknowledgements We thank Timo Rath, Jennifer Krämer, Victoria Dallinger, Joy Einwaller, Shuresh Staudacher, Hanna Rauch, Nikolaus Huber, Ulrike Auer, Eva Eberspächer-Schweda, Carsten Grøndahl, Elisabeth Heiderich, Julia Bohner, who supported the implantation of data loggers and capture events. G. Fluch built the temperature loggers, and we are grateful for his support. We thank Renate Hengsberger for her help in formatting the manuscript. We also thank Erik Pekny for his picture of a wild boar.

Authors contribution CB and TR conceived the ideas. TR led the writing of the manuscript. SV, GS and JP collected the data, and TR analyzed the data. All authors contributed critically to the drafts and gave final approval for publication.

Funding Open Access funding for this article was provided by the University of Veterinary Medicine Vienna (Vetmeduni Vienna). This work was supported by the Austrian Research Promotion Agency (FFG, Grant #855666) and the Governments of Lower Austria and Vienna.

Data availability Upon publication of this article, data will be made available from the University of Vienna PHAIDRA data repository (<https://www.vetmeduni.ac.at/en/bibliothek/infoservice/phaidra/>).

Declarations

Conflict of interest The authors declare no competing interests.

Ethical approval The present study was approved by the Ethics and Animal Welfare Committee of the University of Veterinary Medicine, Vienna, in accordance with the University's guidelines for Good Scientific Practice and authorized by the Austrian Federal Ministry of Education, Science, and Research (ref: BMWFV-68.205/0151-WF/V/3b/2016 and BMWFV-68.205/0224-WF/V/3b/2016) in accordance with current legislation. We confirm that the study was carried out in compliance with the ARRIVE guidelines. No plants or plant parts were used in this study.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are

included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

References

- Baldwin BA, Ingram DL (1968) The influence of hypothalamic temperature and ambient temperature on thermoregulatory mechanisms in the pig. *J Physiol* 198(3):517–529. <https://doi.org/10.1113/jphysiol.1968.sp008622>
- Berthon D, Herpin P, Ledivich J (1994) Shivering thermogenesis in the neonatal pig. *J Therm Biol* 19(6):413–418. [https://doi.org/10.1016/0306-4565\(94\)90040-X](https://doi.org/10.1016/0306-4565(94)90040-X)
- Blix AS (2016) Adaptations to polar life in mammals and birds. *J Exp Biol* 219(8):1093–1105. <https://doi.org/10.1242/jeb.120477>
- Bracke MBM (2011) Review of wallowing in pigs: Description of the behaviour and its motivational basis. *Appl Anim Behav Sci* 132(1):1–13. <https://doi.org/10.1016/j.applanim.2011.01.002>
- Brivio F, Grignolio S, Brogi R, Benazzi M, Bertolucci C, Apollonio M (2017) An analysis of intrinsic and extrinsic factors affecting the activity of a nocturnal species: the wild boar. *Mamm Biol* 84:73–81. <https://doi.org/10.1016/j.mambio.2017.01.007>
- Brown-Brandl TM, Nienaber JA, Xin H, Gates RS (2004) A literature review of pigs heat production. *Trans ASAE* 47(1):259–270. <https://doi.org/10.13031/2013.15867>
- Butler PJ, Green JA, Boyd IL, Speakman JR (2004) Measuring metabolic rate in the field: the pros and cons of the doubly labelled water and heart rate methods. *Funct Ecol* 18(2):168–183. <https://doi.org/10.1111/j.0269-8463.2004.00821.x>
- Chen K, Baxter T, Muir WM, Groenen MA, Schook LB (2007) Genetic resources, genome mapping and evolutionary genomics of the pig (*Sus scrofa*). *Int J Biol Sci* 3(3):153–165. <https://doi.org/10.7150/ijbs.3.153>
- Currie SE, Körtnner G, Geiser F (2014) Heart rate as a predictor of metabolic rate in heterothermic bats. *J Exp Biol* 217(9):1519–1524. <https://doi.org/10.1242/jeb.098970>
- Dexter N (2003) Stochastic models of foot and mouth disease in feral pigs in the Australian semi-arid rangelands. *J Appl Ecol* 40(2):293–306. <https://doi.org/10.1046/j.1365-2664.2003.00792.x>
- Fyda TJ, Spencer C, Jastroch M, Gaudry MJ (2020) Disruption of thermogenic UCP1 predated the divergence of pigs and peccaries. *J Exp Biol* 223(15):jeb223974. <https://doi.org/10.1242/jeb.223974>
- Gaudry MJ, Jastroch M, Treberg JR, Hofreiter M, Paijmans JLA, Starrett J, Wales N, Signore AV, Springer MS, Campbell KL (2017) Inactivation of thermogenic UCP1 as a historical contingency in multiple placental mammal clades. *Sci Adv* 3(7):e1602878. <https://doi.org/10.1126/sciadv.1602878>
- Geisser H, Reyer H-U (2005) The influence of food and temperature on population density of wild boar *Sus scrofa* in the Thurgau (Switzerland). *J Zool (Lond)* 267:89–96. <https://doi.org/10.1017/S095283690500734X>
- Gómez-Prado J, Pereira AMF, Wang D, Villanueva-García D, Domínguez-Oliva A, Mora-Medina P, Hernández-Avalos I, Martínez-Burnes J, Casas-Alvarado A, Olmos-Hernández A, Ramírez-Necochea R, Verduzco-Mendoza A, Hernández A, Torres F, Mota-Rojas D (2022) Thermoregulation mechanisms and perspectives for validating thermal windows in pigs with hypothermia and hyperthermia: An overview. *Front Vet Sci* 9:1023294. <https://doi.org/10.3389/fvets.2022.1023294>
- Green JA (2011) The heart rate method for estimating metabolic rate: Review and recommendations. *Comp Biochem Physiol A* 158(3):287–304. <https://doi.org/10.1016/j.cbpa.2010.09.011>
- Grolemund G, Wickham H (2011) Dates and times made easy with lubridate. *J Stat Softw* 40(3):1–25. <https://doi.org/10.18637/jss.v040.i03>
- Hampton JO, Spencer PBS, Alpers DL, Twigg LE, Woolnough AP, Doust J, Higgs T, Pluske J (2004) Molecular techniques, wildlife management and the importance of genetic population structure and dispersal: a case study with feral pigs. *J Appl Ecol* 41(4):735–743
- Heldmaier G, Neuweiler G, Rössler W (2013) Vergleichende Tierphysiologie. Springer-Lehrbuch, 2., vollständig überarbeitete und aktualisierte Auflage edn. Springer Spektrum, Berlin, Heidelberg
- Jeziński W, Myrcha A (1975) Food requirements of a wild boar population. *Pol Ecol Stud* 1(2):61–83
- Johann F, Handschuh M, Linderoth P, Dormann CF, Arnold J (2020) Adaptation of wild boar (*Sus scrofa*) activity in a human-dominated landscape. *BMC Ecol* 20(1):4. <https://doi.org/10.1186/s12898-019-0271-7>
- Lewis JS, Farnsworth ML, Burdett CL, Theobald DM, Gray M, Miller RS (2017) Biotic and abiotic factors predicting the global distribution and population density of an invasive large mammal. *Sci Rep* 7(1):44152. <https://doi.org/10.1038/srep44152>
- Markov N, Pankova N, Morelle K (2019) Where winter rules: Modeling wild boar distribution in its north-eastern range. *Sci Total Environ* 687:1055–1064. <https://doi.org/10.1016/j.scitotenv.2019.06.157>
- Mellish J-AE, Iverson SJ, Bowen WD (2000) Metabolic compensation during high energy output in fasting, lactating grey seals (*Halichoerus grypus*): metabolic ceilings revisited. *Proc R Soc B* 267:1245–1251. <https://doi.org/10.1098/rspb.2000.1134>
- Muggeo VMR (2003) Estimating regression models with unknown break-points. *Stat Med* 22(19):3055–3071. <https://doi.org/10.1002/sim.1545>
- Nowack J, Giroud S, Arnold W, Ruf T (2017) Muscle non-shivering thermogenesis and its role in the evolution of endothermy. *Front Physiol* 8:889. <https://doi.org/10.3389/fphys.2017.00889>
- Nowack J, Vetter S, Stalder G, Painer J, Kral M, Smith S, Le MH, Jurcevic P, Bieber C, Arnold W, Ruf T (2019) Muscle nonshivering thermogenesis in a feral mammal. *Sci Rep* 9:6378. <https://doi.org/10.1101/377051>
- Olczak K, Nowicki J, Klocek C (2015) Pig behaviour in relation to weather conditions – a review. *Ann Anim Sci* 15(3):601–610. <https://doi.org/10.1515/aoas-2015-0024>
- Phillips PK, Heath JE (1995) Dependency of surface temperature regulation on body size in terrestrial mammals. *J Therm Biol* 20(3):281–289. [https://doi.org/10.1016/0306-4565\(94\)00061-M](https://doi.org/10.1016/0306-4565(94)00061-M)
- Renaudeau D, Leclercq-Smekens M, Herin M (2006) Differences in skin characteristics in European (Large White) and Caribbean (Creole) growing pigs with reference to thermoregulation. *Anim Res* 55(3):209–217. <https://doi.org/10.1051/animres:2006012>
- Ruf T, Vetter SG, Painer J, Stalder G, Bieber C (2021) Atypical for northern ungulates, energy metabolism is lowest during summer in female wild boar (*Sus scrofa*). *Sci Rep* 11(1):18310. <https://doi.org/10.1038/s41598-021-97825-z>
- Ruf T (2022) boxfilter: Filter Noisy Data. 0.1 edn. <https://CRAN.R-project.org/package=boxfilter>
- Thom BT, Huy DTN, Phung TV (2021) Improving yield and quality of wild pork and pigs in Thai Nguyen, Vietnam - and solutions to enhance competitiveness to Thailand and China. *Alinteri J Ag Sci* 36(1):746–752. <https://doi.org/10.47059/alinteri/V36I1/AJAS21104>
- Vetter SG, Ruf T, Bieber C, Arnold W (2015) What is a mild winter? Regional differences in within-species responses to climate

- change. PLoS ONE 10(7):e0132178. <https://doi.org/10.1371/journal.pone.0132178>
- Wehr NH (2021) Historical range expansion and biological changes of *Sus scrofa* corresponding to domestication and feralization. Mamm Res 66(1):1–12. <https://doi.org/10.1007/s13364-020-00534-7>
- Wickham H (2016) ggplot2. Elegant graphics for data analysis. Springer, New York
- Wood SN (2017) Generalized additive models: an introduction with R, 2nd edn. CRC Press, Chapman and Hall, Boca Raton
- Yousef MK (1985) Stress physiology in livestock. Volume I. Basic principles. CRC Press, Boca Raton, Florida

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.