

## RESEARCH ARTICLE OPEN ACCESS

# Taxonomic and Functional Features of Surface to Deep-Sea Prokaryotic Communities in the Eastern North Pacific Ocean

Daniele De Corte<sup>1</sup>  | Leon Dlugosch<sup>2</sup> | Abhishek Srivastava<sup>3,4</sup> | Meinhard Simon<sup>2</sup> | Dennis A. Hansell<sup>5</sup> | Sarah Bercovici<sup>6</sup> | Monica Orellana<sup>7,8</sup>

<sup>1</sup>Ocean Technology and Engineering (OTE), National Oceanography Centre, Southampton, UK | <sup>2</sup>Institute for Chemistry and Biology of the Marine Environment (ICBM), University of Oldenburg, Oldenburg, Germany | <sup>3</sup>Department of Limnology and Bio-Oceanography, University of Vienna, Vienna, Austria | <sup>4</sup>Research Institute of Wildlife Ecology, University of Veterinary Medicine Vienna, Vienna, Austria | <sup>5</sup>Department of Ocean Sciences, University of Miami, Miami, Florida, USA | <sup>6</sup>Ocean BioGeosciences (OBG), National Oceanography Centre, Southampton, UK | <sup>7</sup>University of Washington, Seattle, Washington, USA | <sup>8</sup>Institute for Systems Biology, Seattle, Washington, USA

**Correspondence:** Daniele De Corte ([daniele.decorde@noc.ac.uk](mailto:daniele.decorde@noc.ac.uk)) | Monica Orellana ([morellan@uw.edu](mailto:morellan@uw.edu))

**Received:** 2 June 2025 | **Revised:** 16 July 2025 | **Accepted:** 24 July 2025

**Funding:** This work was supported by Deutsche Forschungsgemeinschaft (CO 2218/2-1, TRR51) and the National Science Foundation (OCE-1634009, OCE-1634250).

**Keywords:** diversity | marine ecosystem | metabolic potential | prokaryotes

## ABSTRACT

Biogeochemical cycles in the ocean are strongly influenced by microbial activity, which affects nutrient and organic matter cycling. These processes, influenced by factors such as temperature, salinity, density and inorganic nutrients, drive the vertical stratification of microbial communities, which subsequently influence the chemistry at different depth layers. Sequencing technology has expanded our understanding of oceanic prokaryotic communities' taxonomic and functional potential. However, there is limited information on how these communities vary across gradients. In this study, we conducted metagenomic analyses on samples from the eastern North Pacific, collected across a longitudinal transect around 45°N and throughout the entire water column. We assessed taxonomic and functional classification, focusing on the roles of prokaryotic communities in biogeochemical cycling. Our results revealed that the surface community was dominated by the SAR11 clade, followed by Flavobacteriales and Rhodobacteriales. The deep layers harboured a more diverse community, where Thaumarchaeota accounted for the most significant proportion. This clear taxonomic stratification led to variations in the communities' functional capabilities across different depth layers. Photosynthesis and heterotrophy dominated the surface layers, whereas the deeper layers exhibited a mix of metabolic features, allowing organisms to potentially utilise both inorganic and organic carbon sources.

## 1 | Introduction

Prokaryotes play a key role in the ocean's biogeochemical cycles by transforming organic matter and recycling nutrients (Azam and Malfatti 2007; Moran 2015). These oceanic crucial biological processes vary greatly across latitude (Dlugosch et al. 2022; Salazar et al. 2019) and pelagic zones (Deutschmann et al. 2024).

Surface processes are dominated by photosynthesis, where inorganic carbon is transformed by phototrophs into organic carbon, making it available to heterotrophic organisms (Karl et al. 1998). The deep ocean mostly relies on surface-derived carbon that sinks into the lower layers as particles and dissolved organic carbon transported via global thermohaline circulation (Aristegui et al. 2009; Herndl and Reinthaler 2013; Carlson et al. 2010).

This is an open access article under the terms of the [Creative Commons Attribution](https://creativecommons.org/licenses/by/4.0/) License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2025 The Author(s). *Environmental Microbiology Reports* published by John Wiley & Sons Ltd.

Several studies have shown that the dark oceans' biogeochemical cycles are not solely dependent on the supply of organic carbon from the euphotic layers. Chemolithoautotrophy and mixotrophy also play critical roles in producing organic carbon through inorganic carbon fixation (Herndl and Reinthaler 2013; Nunoura et al. 2015; Swan et al. 2011). Therefore, the key biogeochemical pathways and players change with depth, following changes in controlling factors such as temperature, salinity, density, inorganic nutrients and organic matter. These factors exert strong selective pressures on microbial communities along the vertical water column, leading to a clear depth stratification of microbial communities in the open ocean (Xue et al. 2020; Liu et al. 2018).

Recent advances in sequencing technology have increased our understanding of the taxonomic and functional potential of prokaryotic communities. Although several studies have investigated the large-scale distribution of prokaryotic communities in the ocean (Dlugosch et al. 2022; Salazar et al. 2019; Sunagawa et al. 2015), little information is available on how their taxonomic composition, functional traits and overall diversity vary across environmental gradients. To expand this information to the understudied temperate North Pacific, we collected samples for metagenomic analyses in the eastern North Pacific at 13 stations on a zonal transect around 45°N, ranging from coastal waters to the open ocean. Samples were collected throughout the water column, from the surface to the bathypelagic layers. The taxonomic diversity was assessed by extracting 16S rRNA gene fragments from the metagenomes (Logares et al. 2014), while the functional features were classified using KEGG orthologues (KOs). The main objectives of this study were (i) to taxonomically and functionally characterise the prokaryotic communities across the transect and different pelagic zones and (ii) to assess the factors controlling the shape of the prokaryotic communities, especially their functional capability.

## 2 | Materials and Methods

### 2.1 | Study Area and Sampling

Sampling was conducted on board the *R/V Oceanus* in August 2018. Water samples were collected at 13 stations through the entire water column along a zonal transect between 124.9°W and 136.0°W. Samples for biological (DNA sequencing) analyses were collected at four to six depths: the surface layer (2–20 m), the subsurface (25–100 m), the mesopelagic (200–450 m), the upper oxygen minimum zone (OMZ) (600 m) and the bathypelagic (2100–4145 m). Water samples were collected with 12-L Niskin bottles mounted on a CTD (conductivity–temperature–depth) rosette sampler equipped with specific pressure, temperature, salinity, conductivity and dissolved oxygen sensors.

### 2.2 | Inorganic Nutrient Concentrations

Samples for dissolved inorganic nutrients (nitrate, nitrite, orthophosphate and silicate) were measured by the Marine Chemistry Lab at the University of Washington, following the protocols of the WOCE Hydrographic Program using a Seal Analytical

AA3 autoanalyzer (SEAL Analytical Inc., Mequon, WI, USA) (UNESCO 1994).

### 2.3 | Dissolved Organic Carbon

DOC was quantified on a Shimadzu TOC-VCSH total organic carbon analyser, equipped with an autosampler ASI-V, via high-temperature catalytic combustion in the Hansell Biogeochemistry Lab at the University of Miami as described in (Halewood et al. 2022). The analyses were quality controlled using a DOC deep-sea reference material (Hansell Biogeochemistry Laboratory, University of Miami; Hansell 2005).

### 2.4 | DNA Extraction and Sequencing

Depending on the depth, 1–5 L of water were filtered onto a 47-mm membrane filter of 0.22 µm pore size (Millipore, GTTP) and stored at –80°C until further processing in the lab. DNA extraction was performed using the DNeasy Power Soil Pro Kit (QIAGEN) following the manufacturer's instructions. The extracted DNA's quality and concentration were assessed by Qubit dsDNA HS Assay Kit (Thermo Fisher Scientific). Illumina shotgun libraries were prepared by the Northwest Genomics Laboratory (University of Washington, WA, USA) using the Kapa Hyper library construction kit (Roche). Sequencing was performed by Illumina NovaSeq 6000 (Illumina Inc., San Diego, USA). The obtained reads were deposited in the NCBI Sequence Read Archive (SRA) under the BioProject accession number PRJNA1002880.

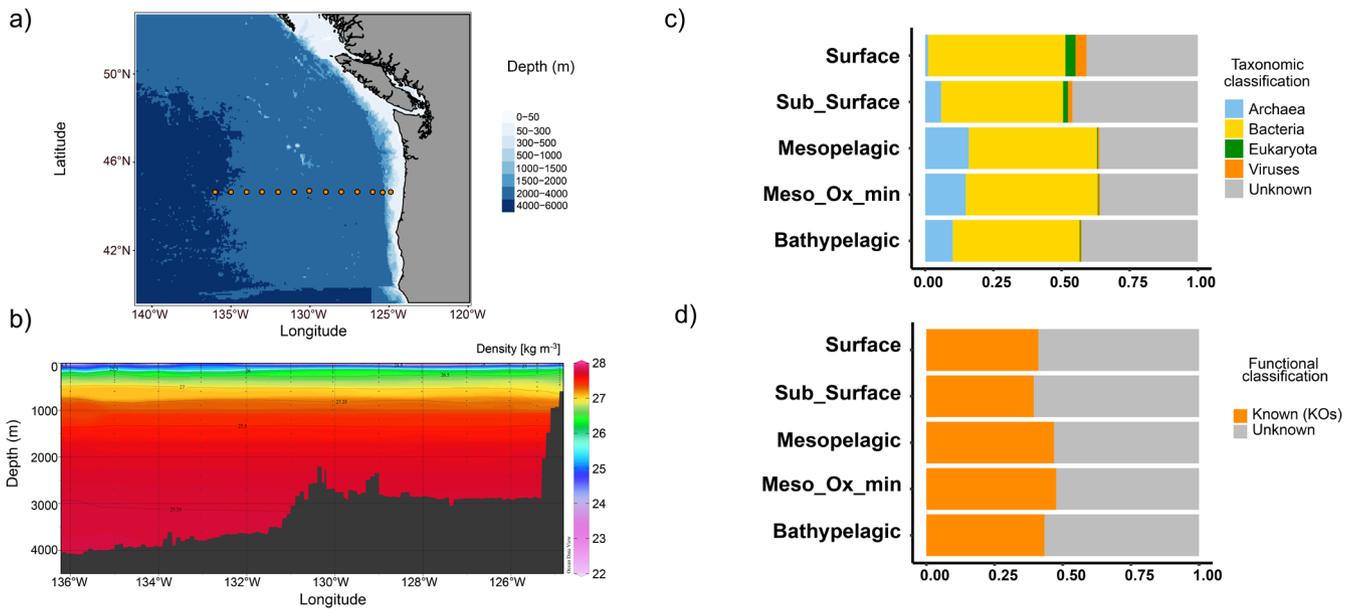
## 3 | Bioinformatics Analyses

### 3.1 | Metagenomic Assembly and Gene Prediction

Reads were quality checked, and adapter sequences were trimmed using Trimmomatic 0.36 (ADAPTER:2:30:10 SLIDINGWINDOW:4:25 MINLEN:100) (Bolger et al. 2014). The high-quality reads were assembled using metaSPAdes 3.11 (Nurk et al. 2017). Coding sequences of the obtained contigs were predicted using Prodigal 2.6.2 (Hyatt et al. 2010). Genes shorter than 210 bp and longer than 4500 bp were discarded. Additionally, the sequences were clustered at 95% identity to generate a gene catalogue using USEARCH v10 (cluster\_fast-id 0.95) (Edgar 2010). The obtained gene sequences were taxonomically classified using Kaiju 1.6 (Menzel et al. 2016) (greedy mode with five allowed substitutions and  $e$ -value  $10e^{-5}$ ) with the Refseq nr (O'Leary et al. 2015) and ProGenomes (Mende et al. 2017) databases and functionally assigned by using the Kyoto Encyclopedia of Genes and Genomes (KEGG) online annotation tool GhostKOALA (Kanehisa et al. 2016) (<https://www.kegg.jp/ghostkoala/>) using the KEGG gene database (release 86) with default settings.

### 3.2 | Read Abundance

Reads longer than 75 bp were mapped to the obtained sequences using bowtie 2.3.5 (very-sensitive-local mode) (Langmead



**FIGURE 1** | (a) Map of the station visited during the cruise in 2018, (b) density plot of whole depth profile and (c) percentages of taxonomic (Bacteria, Viruses, Archaea, Eukaryota and unclassified) and functional (KEGG and unclassified) annotations of gene sequences of the metagenomes.

and Salzberg 2012). SAMtools version 1.9-58-gbd1a409 (Li et al. 2009) was used to produce the read abundance table from the alignment file. The obtained data was normalised by dividing read counts by the length of each gene in kb to obtain reads per kilobase (RPK). Subsequently, the sum of RPKs for each sample was divided by  $10^6$  to obtain reads per kilobase per million (RPKM).

### 3.3 | Metagenomics 16S rRNA Gene Tags

16S ribosomal RNA gene fragments were extracted from the metagenome reads as described by Logares et al. (2014). The extracted 16S miTags were further processed using USEARCH v11 (Edgar 2013, 2018). Briefly, the obtained miTags were clustered at 97% sequence identity, and a counts table was constructed by mapping the 16S reads to the final OTUs. Finally, the OTUs were taxonomically assigned using SINTAX (Edgar 2016) with the RDP 16S training set v16 database (Maidak et al. 2001). OTUs reported in more than 80% of the samples with detection of at least 0.1% were considered core phylotypes.

### 3.4 | Data Processing

All data processing and statistical analyses were conducted in R software v.3.6 (version 3.6.0; <https://www.r-project.org/>) with the following additional packages: vegan (Oksanen et al. 2018), pheatmap (Kolde 2019), DESeq2 (Love et al. 2014), phyloseq (McMurdie and Holmes 2013), microbiome (Lahti and Shetty 2019) and ggOceanMaps (Vihtakari 2024). Linear interpolation was used to estimate missing nutrient values ( $n=1$ ) based on known values within the data set. Ocean Data View (ODV) (Schlitzer 2023) was used to calculate oxygen saturation and generate the density plot shown in Figure 1.

## 4 | Results

### 4.1 | Study Area Physicochemical Characteristics

There was a surface-to-depth stratification of water masses, with density values decreasing from a minimum of  $22.5 \text{ kg m}^{-3}$  at the surface to a maximum of  $27.8 \text{ kg m}^{-3}$  in the bathypelagic layers. Dissolved oxygen concentrations decreased with depth from the highest values of  $271.9\text{--}291.9 \mu\text{mol kg}^{-1}$  (oxygen saturation 99.3%–105.7%) in surface and subsurface waters to  $14.3\text{--}28.8 \mu\text{mol kg}^{-1}$  (oxygen saturation 4.6%–9.1%) at 600 m depth (Table 1). The oxygen concentrations below the mesopelagic oxygen minimum (OMZ) (600 m) increased towards the bathypelagic layers, ranging between  $60.7$  and  $121.6 \mu\text{mol kg}^{-1}$  (oxygen saturation 18.1%–35.7%). Nitrate and phosphate concentrations increased with depth, reaching a maximum within the OMZ ( $43.5$  and  $3.19 \mu\text{mol kg}^{-1}$ , respectively). Nitrite concentrations showed no trend among different depth layers (Table 1). Silicate increased with depth, with the lowest values detected at the surface ( $1.0 \mu\text{mol kg}^{-1}$ ) and the highest ( $162.7 \mu\text{mol kg}^{-1}$ ) in the bathypelagic layers. On the other hand, DOC showed an opposite trend, with its value declining from  $78.1$  at the surface to  $36.4 \mu\text{mol kg}^{-1}$  in the bathypelagic layers (Table 1).

### 4.2 | Microbial Community Structure

After assembly, a total of  $6.3 \times 10^6$  unique nr protein-coding genes were predicted from the microbial community. About 44% of these coding genes were functionally annotated by homology to KEGG ORTHOLOGY (KO) (Figure 1d). The largest proportion of genes was affiliated with Bacteria (~47%), followed by Archaea (~10%), Eukaryotes (~1%) and Viruses (~1%) (Figure 1c). The Archaea contribution increased with depth and reached its maximum value in the mesopelagic

**TABLE 1** | Physico chemical characteristics of the sampled waters in the Eastern Nord Pacific.

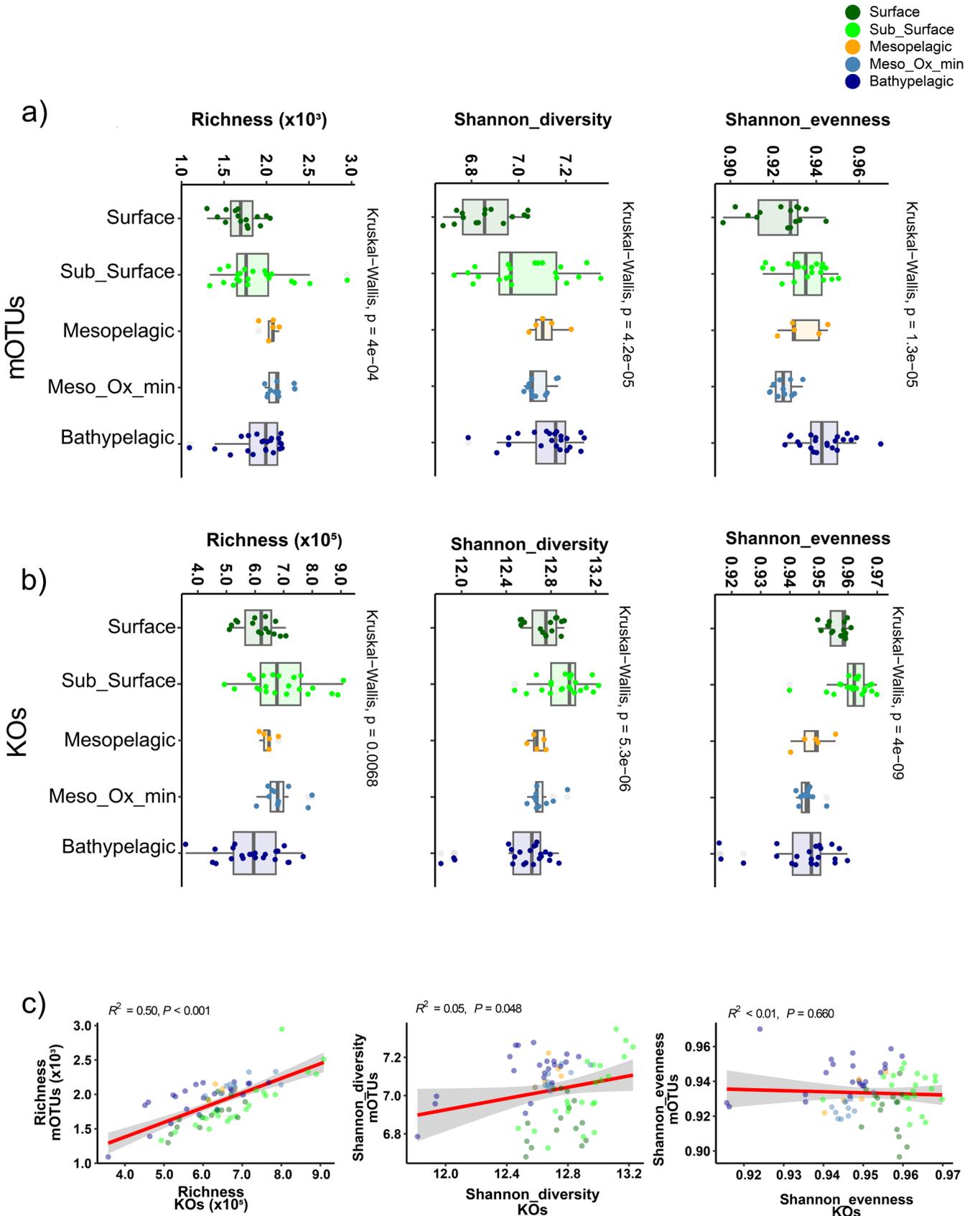
	No. of samples	Depth (m)	Temperature (°C)		Salinity		Density (kg m <sup>-3</sup> )	Oxygen		Si (μmol kg <sup>-1</sup> )	NO <sub>3</sub> <sup>-</sup> (μmol kg <sup>-1</sup> )	NO <sub>2</sub> <sup>-</sup> (μmol kg <sup>-1</sup> )	PO <sub>4</sub> <sup>3-</sup> (μmol kg <sup>-1</sup> )	DOC (μmol kg <sup>-1</sup> )
			Depth	Temperature	Salinity	Oxygen		Oxygen Sat (%)						
Surface	15	2–20	10.5–18.5	31.5–32.7	22.5–24.9	227.2–271.9	95.8–99.3	1.0–9.6	0.03–1.99	0.00–0.10	0.10–0.57	59.3–78.1		
Subsurface	22	25–100	7.5–14.8	32.4–33.7	24.2–26.1	132.7–291.9	46.6–105.7	1.7–28.0	0.09–25.13	0.01–0.54	0.39–1.82	48.2–67.4		
Mesopelagic	5	200–450	7.8–5.7	33.9–34.1	26.5–26.9	43.7–147.2	14.3–50.5	37.9–64.8	28.25–39.14	0.01–0.03	1.91–2.76	41.9–52.2		
Meso Ox-min	11	600	4.2–5.0	34.1–34.2	27.0–27.1	14.3–28.8	4.6–9.1	51.7–91.2	22.9–43.5	0.00–0.02	2.06–3.19	39.6–48.8		
Bathypelagic	22	2100–4145	1.5–1.9	34.6–34.7	27.7–27.8	60.7–121.6	18.1–35.7	92.7–162.7	21.6–42.5	0.00–0.09	1.87–2.95	36.4–46.8		

layers, while the contribution of eukaryotes and viruses to the total microbial community declined with depth (Figure 1c). The 16S rRNA gene OTUs' taxonomic and KOs profiles varied significantly with depth in richness, diversity and evenness (Figure 2). Taxonomic and KOs richness were correlated ( $R^2 = 0.50$ ,  $p < 0.001$ ), while diversity and evenness showed different patterns between the two profiles. The taxonomic profiles displayed increased diversity and evenness from the surface to the bathypelagic layers, whereas those of the KOs profiles decreased. This resulted in a minimal relation between the two profiles in terms of Shannon diversity and evenness (Figure 2). The PCA of the metagenome extracted OTUs (mOTUs) and KOs profiles showed that the microbial communities were stratified following the changes in water density. PC1 and PC2 accounted for 31.5% and 6.4% of the variation in the mOTUs profiles, while the KOs PC1 and PC2 accounted for 36.7% and 12.5%, respectively. Variation partitioning showed that sample distance and physicochemical variables explained ~40% of the variation of the mOTUs profile and ~50% of the KOs profile, with physico-chemical variables accounting for most of the variation in both profiles (Figure 3b,d).

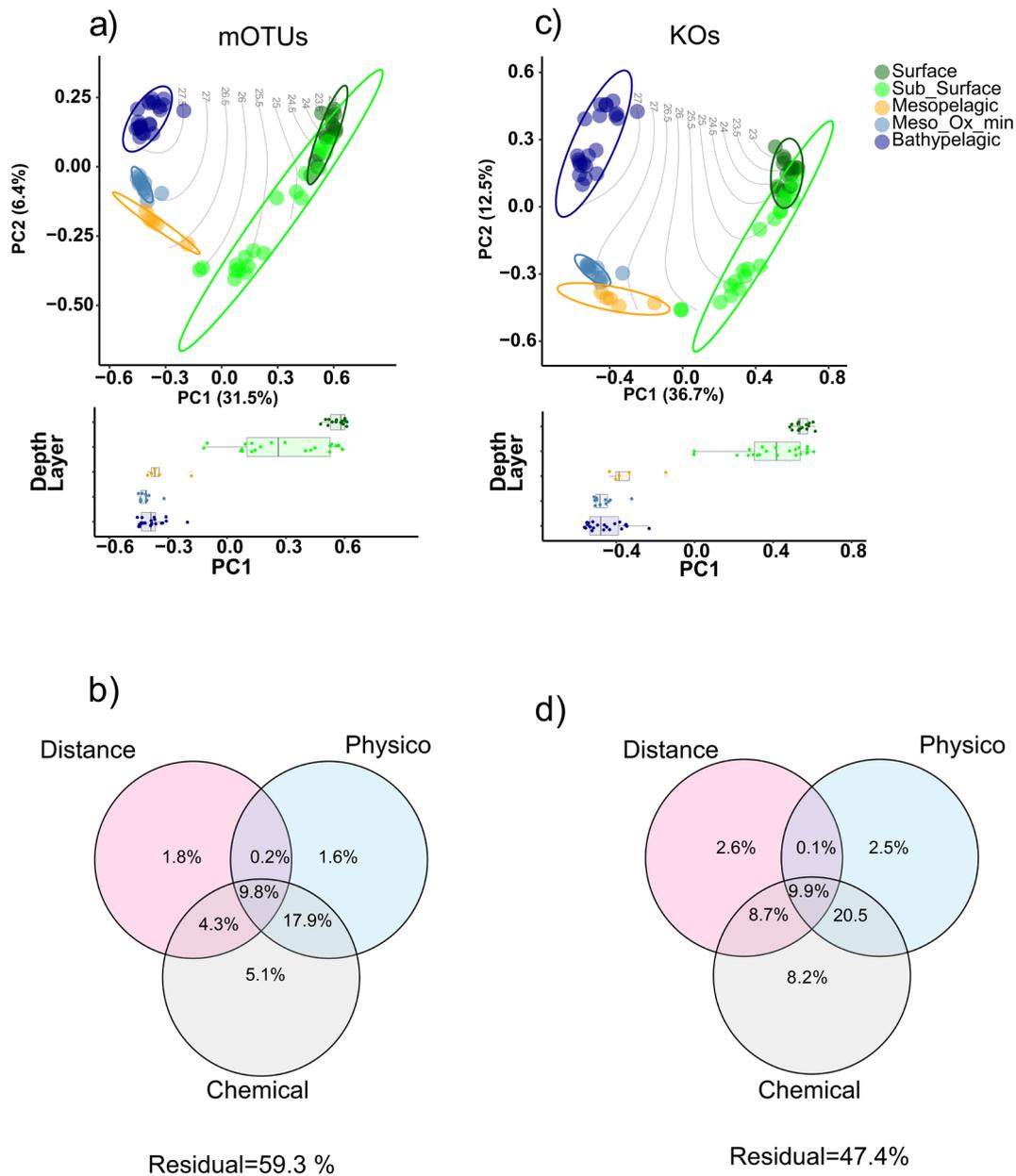
The taxonomic and functional profiles were grouped along the water column following the depth and density gradient as shown by the Bray–Curtis similarity matrices (Figure 4a,c). The relationships between density distance and the similarities of the microbial community of the mOTUs and KOs showed similar distribution, with the similarity of the microbial community declining with the increase in density differences. Samples collected at the same density (i.e., where the density distance is 0) showed a microbial similarity ranging between 20% and 70%, suggesting high microbial community variability within the same water mass. On the other hand, samples with high-density distance showed very low similarity in microbial community composition, especially in the KOs profile (Figure 4b,d). Additionally, the KOs and mOTUs similarity profiles were highly correlated ( $R^2 = 0.88$ ;  $p < 0.001$ ; Figure 4e).

### 4.3 | Taxonomic Structure of the Microbial Communities

Phylogenetic analyses of the extracted 16S rRNA genes (mOTUs) and the metagenome gene sequences (KOs) revealed the dominance of *Proteobacteria*, *Bacteroidetes*, *Thaumarchaeota* and *Actinobacteria* (Figure 5a,b). The microbial communities showed high taxonomic variability in both profiles, with clear changes in depth, with the KOs profiles showing higher resolution in the number of taxa found than the mOTUs. In contrast, microbial communities at the surface did not change from the near shore to the open ocean. The *Pelagibacteriales* largely contributed to the microbial communities in both profiles; their contribution was larger in surface (reaching 33% in the KOs and 50% in the mOTUs profiles) and declined with depth, reaching their lowest values in the bathypelagic layers: ~15% in the KOs profiles and 24% in the mOTUs. *Flavobacteriales* and *Rhodobacteriales* also significantly contributed to the microbial communities in both profiles in the surface and subsurface layers. Their contribution



**FIGURE 2** | Richness, Diversity and evenness indexes of mOTUs (a) and KOs (b) profiles across five different pelagic zones. Relation between the mOTUs and KOs indexes (c).



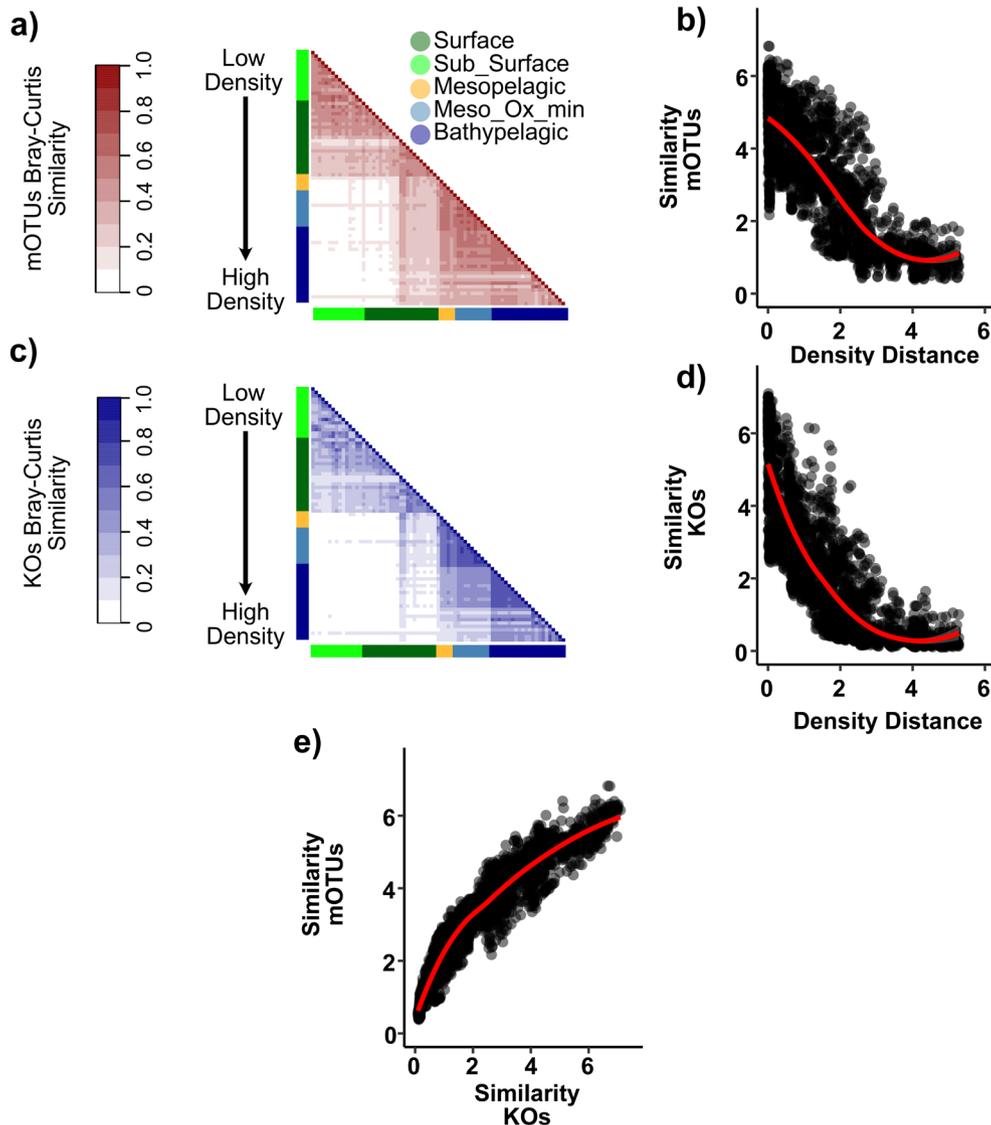
**FIGURE 3** | Principal component analyses (PCAs) of mOTUs (a) and KO profiles (c). Variation partitioning of physico-chemical and spatial factors explaining the prokaryotic community composition of the taxonomic (b) and functional (d) profiles.

declined with depth and reached lower values in the bathypelagic layers. In the mOTUs profile, *Nitrosopumilaceae* had the largest contribution in the mesopelagic and bathypelagic layers, reaching ~30% of the total communities. In the KOs profiles, unidentified Euryarchaeota and Thaumarchaeota also largely contributed to the meso- and bathypelagic community (Figure 5a,b).

In the taxonomic profile, 39 OTUs were assigned to *Candidatus* Pelagibacter, and 4 were putatively assigned to Unclassified Alphaproteobacteria, which composed the core surface microbial community. Ninety-one taxa formed the mesopelagic core community. The most abundant taxa of the mesopelagic core community were assigned to *Candidatus* Pelagibacter (18%), *Nitrosopumilus* (16%), followed by unassigned Bacteria (16%), Proteobacteria (14%), Gammaproteobacteria (14%) and Thaumarchaeota (10%). The bathypelagic core community

was composed of 53 taxa dominated by mOTUs with similar taxonomic affiliations as those found in the mesopelagic core community; ~92% of the bathypelagic core community was shared with the mesopelagic community. All three depth layers shared nine core taxa identified as *Candidatus* Pelagibacter (Figure S1).

Among the potential nitrifiers, apart from *Nitrosopumilaceae* that dominated the meso- and bathypelagic zones, *Nitrosomonadales* (which includes five genera of ammonia-oxidising bacteria) were also found in the surface (~1%) layers. (*Nitrosomonadales* abundance is only displayed in the KOs profile as in the mOTUs profile their median across the samples was below 0.1%). Nitrite-oxidising bacteria associated with the *Nitrospinales* order were also detected in both profiles, with their contribution maximised in the OMZ and mesopelagic layers.



**FIGURE 4** | Similarity matrices of mOTUs (a) and KOs (c) profiles and their changes in comparison with density (b, d). Relation between the mOTUs and KOs Bray–Curtis similarities (e).

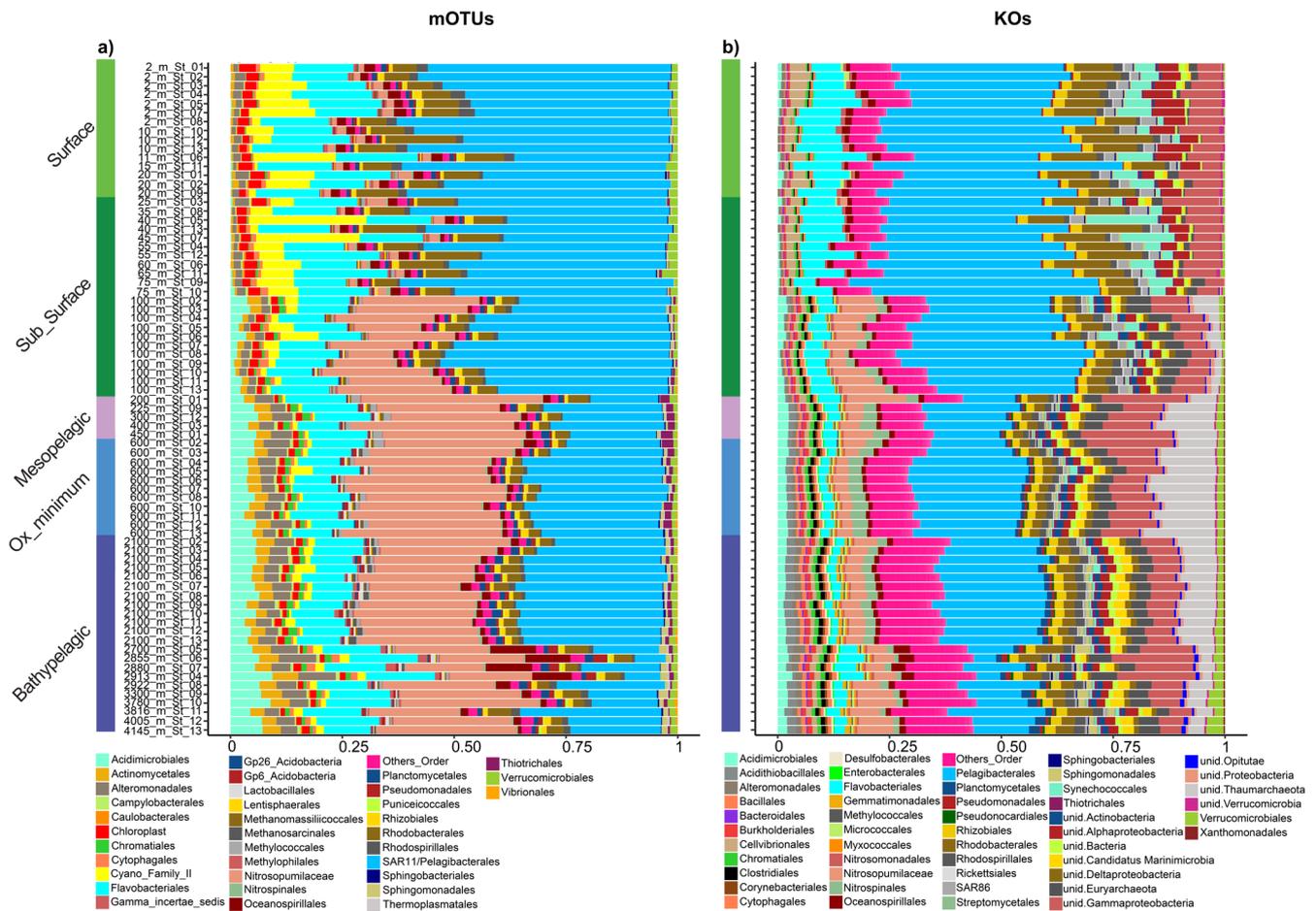
#### 4.4 | Functional Analysis of Microbial Community

The metagenomic data obtained from the microbial communities were used to characterise potential metabolic pathways. The KOs showed a clear depth-dependent pattern, with most assigned genes clustering within different depth layers (Figure S2). To explore the potential metabolic differences between microbial communities inhabiting different pelagic zones, marker genes related to relevant pathways in the open ocean were selected (Figures 6 and 7). These marker genes were characterised by their functions in phototrophy, carbon fixation, CO-oxidation, respiration, nitrogen, sulphur and methane-phosphonate metabolisms.

The surface community was dominated by phototrophic mechanisms, along with genes encoding for carbon dioxide and nitrogen fixation. Carbon monoxide oxidation genes were significantly higher in the meso- and bathypelagic layers compared to the surface ocean (Figures 6 and 7). This process involves converting carbon monoxide into CO<sub>2</sub>, using

oxygen as the primary electron acceptor, or alternative acceptors like nitrate or sulphate in low-oxygen conditions (King and Weber 2007). Other processes that occur where oxygen is limited or absent, such as denitrification and dissimilatory nitrate reduction, were up to twofold higher in the meso- and bathypelagic layers than in the surface ocean. On the other hand, assimilatory nitrate reduction, a process that converts nitrate (NO<sub>3</sub><sup>-</sup>) to ammonium (NH<sub>4</sub><sup>+</sup>), which is then incorporated into organic molecules, dominated in the epipelagic layers. This process is particularly important for the uptake of organic nitrogen by primary producers (both eukaryotes and cyanobacteria).

While genes associated with sulphur metabolism did not show clear differences between depth layers, thiosulfate oxidation was primarily found in the surface ocean (Figures 6 and 7). Genes associated with methanogenesis did not show a clear trend through the water column. In contrast, coenzyme M biosynthesis, another gene involved in the methane cycle, was predominant in the surface ocean. Methylphosphonate



**FIGURE 5** | Taxonomic classification of the mOTUs (a) and KOs (b) profiles across the longitudinal transect and the water column.

metabolism genes, involved in the degradation of organic matter that serves as a source of phosphorus, had a higher contribution in the surface and bathypelagic layers. In contrast, their contribution was minimal in the mesopelagic layers (Figures 6 and 7).

## 5 | Discussion

### 5.1 | Factors Driving Prokaryotic Taxonomic and Functional Composition in the Water Column

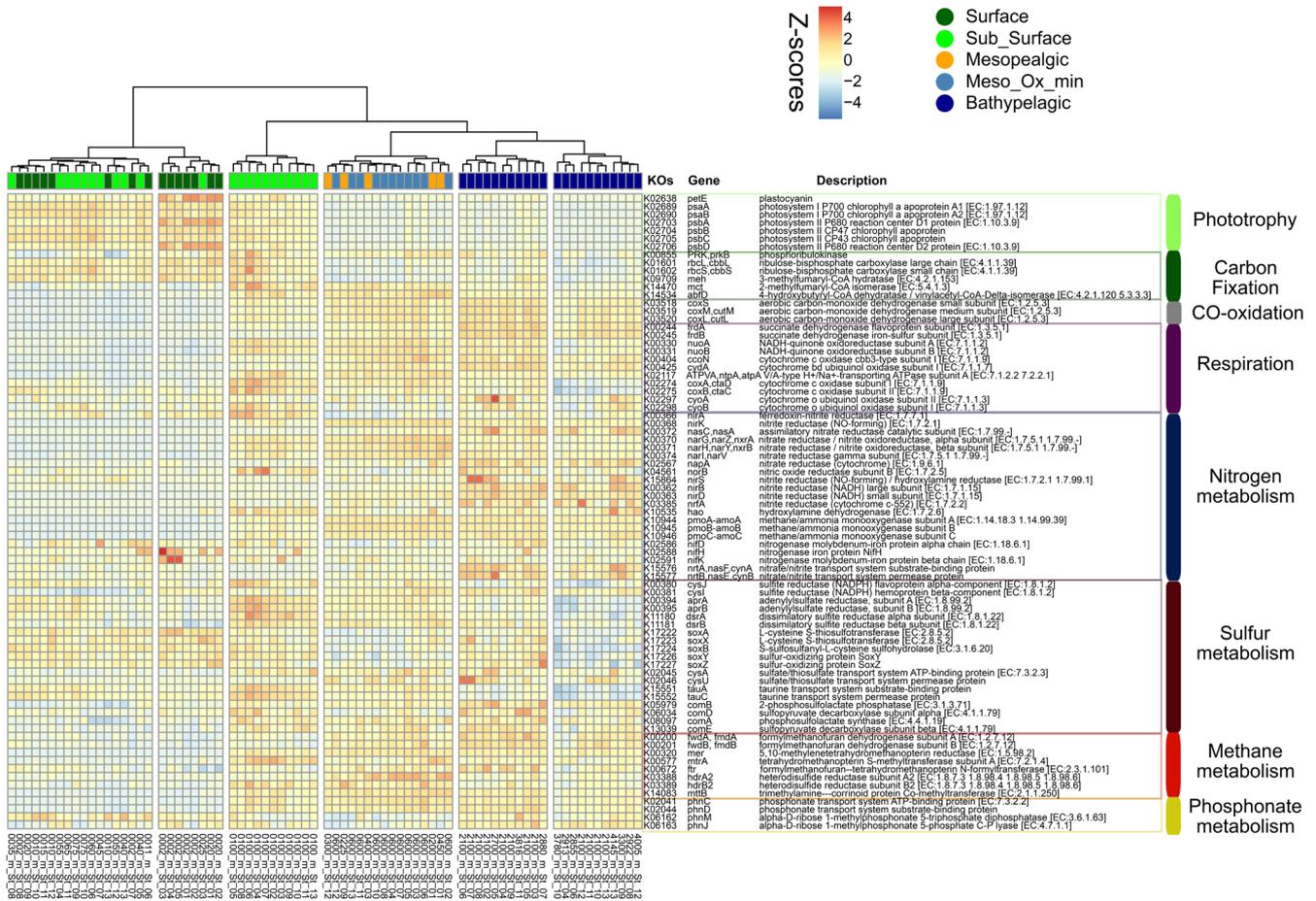
Understanding the factors that shape prokaryotic taxonomic and functional composition throughout the water column is essential for interpreting microbial biogeography and ecosystem functioning. Surface-to-deep ocean profiles across a transect provide important insights into vertical and regional patterns. Such comprehensive data sets are vital for achieving broader spatial coverage and uncovering region-specific drivers of microbial community structure and function.

The mOTUs' richness and diversity increased from surface to the meso- and bathypelagic layers, while the KOs indexes appeared lower in the deeper layer, suggesting a reduced functionality of the prokaryotic community in the dark ocean within a broader community. The dark ocean harbours a wide range of taxonomically diverse organisms that survive under

limited carbon sources (Arrieta et al. 2015). In fact, mOTU diversity had a negative relationship with DOC ( $R = -0.61$ ,  $p < 0.001$ ) (Figure S3), suggesting that the microbial community increases its diversity as the concentration of available substrate (measured here as DOC) decreases. Both chemical recalcitrance of individual molecules and their individual diluted concentrations play a role in limiting microbial utilisation of DOC as a substrate (Arrieta et al. 2015; Bercovici et al. 2021; Shen and Benner 2018).

Deep-sea microbes rely primarily on labile dissolved organic matter derived from sinking particles from surface waters as a source of growth (Aristegui et al. 2009; Bercovici et al. 2021; Moran et al. 2016; Hansman et al. 2009). While bacterial diversity (Shannon diversity) is high at depth, there are localised areas where it decreases at the seafloor and reflects the Shannon diversity values in the surface waters; this could be due to the solubilisation of sinking particles or resuspension of sediment organic matter (Figure S4b,c).

The surface ocean appears to have different controls on its microbial community. Where the surface DOC concentration is highest, mOTU diversity is highest on the surface (upper ~100 m; Figure S4). This high concentration of available substrates in the surface ocean increases microbial diversity and provides more opportunities for a wider variety of microbial communities to grow.



**FIGURE 6** | Heatmap of selected genes (KOs) accounting for phototrophy, carbon fixation, CO-oxidation, respiration, nitrogen, sulphur, methane, and phosphonate pathways across the longitudinal transect and the water column.

The functional similarity of the prokaryotic community in waters with identical density was higher than taxonomic similarity (Figure 4). This outcome suggests that substrate composition may be selected by different communities that occupy the same ecological niche, indicating that different taxa perform the same ecological function. This redundancy in ecological roles suggests that the communities encode for similar metabolic genes, reflecting redundancy in metabolic capability. Under limiting and stable environmental conditions, closely related species may occupy different ecological niches. While this can lead to high taxonomic diversity, it may not translate into increased functional diversity if specific groups perform similar ecological roles despite their taxonomic differences. Although there may also be adaptation differences among genes associated with a single KO due to varying environmental conditions, this may suggest high niche partitioning among gene variants, as demonstrated by Dlugosch et al. (2022). Our results support the notion that environmental conditions play a key role in shaping the distribution of functional groups while also affecting the taxonomic composition (Louca et al. 2016).

The taxonomic and functional profiles revealed clear differences in bacterial communities across depth (Figures 3 and 4). These results showed that vertical stratification driven by changes in physico-chemical variables over the different depth layers is a key factor in shaping bacterial community structure in the water column. Our correlation analysis provides new insights

by highlighting specific associations between environmental variables and community composition, contributing to a more detailed understanding of these relationships. Subsurface (20–100 m) layers exhibited high richness and diversity in both profiles, indicating that this zone is a hotspot for microbial activity driven by environmental factors and resource availability (Figure 2). This zone, which includes the deep chlorophyll maximum, is characterised by high phytoplankton biomass and production (Cullen 2015), which can promote complex microbial interactions that influence prokaryotic productivity and diversity (Sunagawa et al. 2015; Marañón et al. 2021; Hu et al. 2022).

## 5.2 | Taxonomic Profiling of the Prokaryotic Community

The taxonomic classification of the prokaryotic community along the transect and through the entire water column was conducted using metagenome-extracted 16S rRNA gene (mOTUs) and gene data (KOs). In general, the two profiles were comparable. Nevertheless, the KOs data showed higher taxonomic resolution than the 16S rRNA gene sequences at all taxonomic levels, as previously found in different studies (Durazzi et al. 2021; Khachatryan et al. 2020). Specific orders of prokaryotes showed different distribution patterns across various depths (Figure 6). The SAR11 clade, as previously described (Vergin et al. 2013; Salter et al. 2015; Ortmann and Santos 2016;

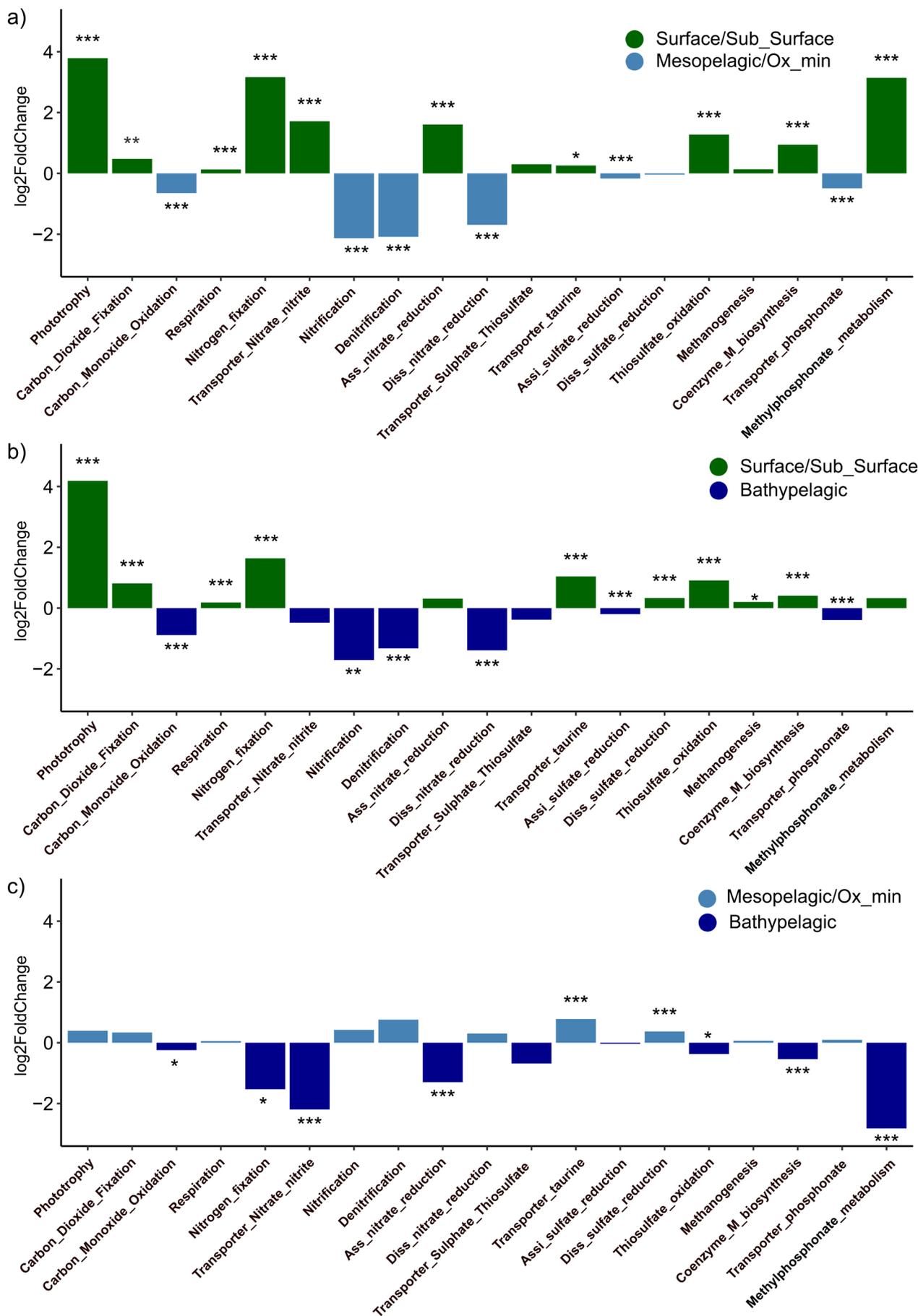


FIGURE 7 | Legend on next page.

**FIGURE 7** | Changes of the selected metabolic pathways between surface/subsurface, mesopelagic/oxygen-minimum and bathypelagic layers. (\*\* $p < 0.001$ ; \*\* $p < 0.01$ ; \* $p < 0.05$ ).

Morris et al. 2002; Milke et al. 2022), dominated the surface and subsurface prokaryotic community in both profiles, with mOTUs taxonomically assigned as *Candidatus Pelagibacter* that constituted 90% of the core surface community. The physiological adaptability traits (such as efficient energy acquisition strategies) of members of the SAR11 clade make this group one of the most abundant members of the open water prokaryotic community that greatly contribute to the ocean's biogeochemical cycles (Giovannoni 2017). Nevertheless, its contribution was not only limited to the surface ocean but also represents a large proportion of the prokaryotic community in the meso- and bathypelagic layers. Several SAR11 genotypes are specialised for living under specific environmental conditions (López-Pérez et al. 2020). A deep SAR11 ecotype, which contains genomic adaptations to the deep ocean and is phylogenetically distinct from the surface clades, was discovered in mesopelagic waters (Thrash et al. 2014). These findings suggest that different SAR11 types occupy different ecological niches in different pelagic zones.

*Flavobacteriales* and *Rhodobacterales* also contributed to the surface and subsurface communities, and their abundance peaked where the DOC concentration was higher (Figure S4). These groups play an important role in phytoplankton–prokaryotic interactions by metabolising organic matter of phytoplankton origin (Buchan et al. 2014; Luria et al. 2017). The hydrolytic activity of *Flavobacteriales* on high molecular weight DOM and in particular polysaccharides (Reintjes et al. 2019) may promote the release of low molecular weight organic matter that can be used by other members of the prokaryotic community, such as *Rhodobacterales* and SAR11.

Orders associated with Archaea showed very low abundance in the euphotic layers. Their contribution significantly increased and reached its largest proportion in the meso- and bathypelagic layers, where they were also a main member of the core prokaryotic community together with other groups. The lower abundance in the upper euphotic zone is consistent with previous studies showing the high sensitivity of ammonia-oxidising Archaea to photoinhibition (Nunoura et al. 2015; Merbt et al. 2012). Many marine Thaumarchaeota, such as *Nitrosopumilus*, are involved in the oxidation of ammonia, a crucial step in the nitrogen cycle. This metabolic capability allows them to exploit a niche where they can use ammonia as an alternative source of energy when bioavailable organic matter is limited, as it often is in the meso- and bathypelagic ocean (Wright and Lehtovirta-Morley 2023; Sintes et al. 2013). Ammonia-oxidising Bacteria, such as *Nitrosomonas* and *Nitrosospira*, were predominantly found in the surface layers, indicating a niche separation between ammonia-oxidising Bacteria and Archaea in different pelagic zones (Nunoura et al. 2015).

Additionally, the co-occurrence of ammonia-oxidising Archaea and nitrite-oxidising Bacteria below the euphotic layers suggests a complementary relationship in the nitrification process, where the spatial and functional niche differentiation across various

pelagic zones ensures an efficient nitrogen cycle in which each group specialises in different stages of the process.

### 5.3 | Functional Characterisation of the Prokaryotic Community

Chlorophyll-subunit-encoding genes were primarily detected in the surface and sub-surface ocean layers. However, they were also present in lower abundance in deeper layers, suggesting that sinking particles of photosynthetic origin are actively transported to the ocean's interior and may serve as additional bioavailable organic carbon (through particle solubilisation) for deep heterotrophic marine microbes (Dang et al. 2024; Agusti et al. 2015; Nagata et al. 2000; Hansell and Ducklow 2003). In our study, carbon fixation was associated with diverse microbial taxa. Sunlight-driven primary productivity was linked to ribulose-1,5-bisphosphate carboxylase (RuBisCo) subunit genes. Interestingly, genes encoding both the small and large subunits of RuBisCo were also found in the dark ocean (Figure 6), in agreement with earlier studies that reported RuBisCo proteins (Orellana and Hansell 2012) and high rates of inorganic carbon fixation in bathypelagic waters (Reintaler et al. 2010). These findings suggest that carbon fixation via RuBisCo is not limited to the surface ocean but is a widespread process that may also occur via chemolithoautotrophy in the dark (Swan et al. 2011; De Corte et al. 2021). It is also possible that a portion of the RuBisCo-related genes detected in deep waters originate from phytoplankton transported by sinking particles from the photic zone (Orellana and Hansell 2012). The mesopelagic layers also contained genes for the 3-Hydroxypropionate/4-Hydroxybutyrate (HP/HB) cycle, specifically encoding 4-hydroxybutyryl-CoA dehydratase (4HBD), which were predominantly linked to Thaumarchaeota. The presence of ammonia monooxygenase-encoding genes (*amoA*, *amoB* and *amoC*) in the mesopelagic zone further suggests a link between the HP/HB cycle and carbon fixation in archaeal members of the *Nitrosopumilaceae* family, as previously reported (Könneke et al. 2014). Carbon monoxide dehydrogenase (CODH), encoded by the genes *coxL*, *coxM* and *coxS*, mediates the aerobic oxidation of CO. The increased abundance of these genes in the mesopelagic and bathypelagic layers is consistent with earlier findings (Martín-Cuadrado et al. 2007) and suggests that deep-sea microbes may oxidise carbon monoxide as an alternative energy source in environments where energy and bioavailable organic matter are scarce or only sporadically available, such as the mesopelagic and deep ocean.

The co-occurrence of sulphur oxidation genes alongside Calvin cycle-related genes in the surface and subsurface layers suggests that thiosulfate may be utilised as an electron donor in microbial energy metabolism, driving sulphur oxidation processes as previously reported in the deep ocean (Srivastava et al. 2023). This metabolic strategy allows these microbes to simultaneously oxidise thiosulfate and fix carbon through the Calvin cycle, providing energy to sustain growth while avoiding competition

with heterotrophs. Additionally, CO<sub>2</sub> reduction/fixation genes, such as formyl-methanofuran dehydrogenase subunit-encoding genes (*fwdA* and *fwdB*), were more abundant in meso- and bathypelagic layers than in surface waters. This finding suggests that the reductive acetyl-CoA pathway may function in tandem with methane metabolism among Euryarchaeota, such as *Methanosarcinales* (Figures 5a and 6) (Schöne et al. 2022). Finally, our data confirmed the global presence of phosphonate-related genes throughout the entire water column (Lockwood et al. 2022), with a higher prevalence in the bathypelagic and surface ocean (Figure 7).

## 6 | Conclusion

The microbial communities were well stratified across different depths, where different microbial groups occupy and thrive in distinct ecological niches based on their metabolic capabilities and resource requirements. The meso- and bathypelagic ocean layers are environments with limited resources, where different microbial strategies are employed to utilise pulsed and short-term inputs of various energy sources and substrates. Both chemolithoautotrophy (where organisms obtain energy from inorganic chemicals) and heterotrophy (where organisms rely on organic compounds from other organisms) play significant roles. However, our findings also highlight the importance of mixed metabolic strategies where organisms utilise both inorganic and organic carbon sources. These findings highlight the importance of obtaining depth-resolved profiles from a broader range of oceanic regions, including those with OMZs, to better understand spatial variability in microbial community structure and function and to uncover potentially novel metabolic processes.

Finally, our data showed that even when genetic sequences are classified into known taxonomic groups, their functions may still be unknown or poorly understood. Most of the genes found in our metagenomes were not taxonomically or functionally classified, indicating that key components of the ocean's prokaryotes remain unknown.

### Author Contributions

**Daniele De Corte:** conceptualization, investigation, validation, methodology, writing – review and editing, funding acquisition, writing – original draft, visualization, formal analysis. **Leon Dlugosch:** writing – review and editing, methodology, validation, visualization, data curation, formal analysis. **Abhishek Srivastava:** writing – review and editing, methodology, visualization, formal analysis. **Meinhard Simon:** writing – review and editing, funding acquisition. **Dennis A. Hansell:** writing – review and editing, funding acquisition. **Sarah Bercovici:** writing – review and editing, visualization, methodology. **Monica Orellana:** writing – review and editing, funding acquisition.

### Acknowledgements

We thank the captain and crew of *R/V Oceanus* for their support and splendour atmosphere on board. This work was supported by NSF OCE-1634009 and OCE-1634250 to Monica Orellana and Dennis A. Hansell within the collaborative research project “DOC removal in the ocean according to polymer gel theory” and by Deutsche Forschungsgemeinschaft (DFG) within the Transregional Collaborative

Research Center Roseobacter TRR51 to Meinhard Simon. Daniele De Corte was supported by the Deutsche Forschungsgemeinschaft (DFG) projects CO 2218/2-1 (PN: 445462226).

### Conflicts of Interest

The authors declare no conflicts of interest.

### Data Availability Statement

Sequence data are available at the NCBI Sequence Read Archive (SRA) under the BioProject accession number PRJNA1002880 (<https://www.ncbi.nlm.nih.gov/bioproject/?term=PRJNA1002880>). All data described are available upon request from the corresponding author.

### References

- Agusti, S., J. I. González-Gordillo, D. Vaqué, et al. 2015. “Ubiquitous Healthy Diatoms in the Deep Sea Confirm Deep Carbon Injection by the Biological Pump.” *Nature Communications* 6: 7608.
- Aristegui, J., J. M. Gasol, C. M. Duarte, and G. J. Herndl. 2009. “Microbial Oceanography of the Dark Ocean's Pelagic Realm.” *Limnology and Oceanography* 54, no. 5: 1501–1529.
- Arrieta, J. M., E. Mayol, R. L. Hansman, G. J. Herndl, T. Dittmar, and C. M. Duarte. 2015. “Ocean Chemistry. Dilution Limits Dissolved Organic Carbon Utilization in the Deep Ocean.” *Science* 348, no. 6232: 331–333.
- Azam, F., and F. Malfatti. 2007. “Microbial Structuring of Marine Ecosystems.” *Nature Reviews. Microbiology* 5, no. 10: 782–791.
- Bercovici, S. K., M. C. Arroyo, D. de Corte, T. Yokokawa, and D. A. Hansell. 2021. “Limited Utilization of Extracted Dissolved Organic Matter by Prokaryotic Communities From the Subtropical North Atlantic.” *Limnology and Oceanography* 66, no. 6: 2509–2520.
- Bolger, A. M., M. Lohse, and B. Usadel. 2014. “Trimmomatic: A Flexible Trimmer for Illumina Sequence Data.” *Bioinformatics* 30, no. 15: 2114–2120.
- Buchan, A., G. R. LeClerc, C. A. Gulvik, and J. M. González. 2014. “Master Recyclers: Features and Functions of Bacteria Associated With Phytoplankton Blooms.” *Nature Reviews Microbiology* 12, no. 10: 686–698.
- Carlson, C. A., D. A. Hansell, N. B. Nelson, et al. 2010. “Dissolved Organic Carbon Export and Subsequent Remineralization in the Mesopelagic and Bathypelagic Realms of the North Atlantic Basin.” *Deep Sea Research Part II: Topical Studies in Oceanography* 57, no. 16: 1433–1445.
- Cullen, J. J. 2015. “Subsurface Chlorophyll Maximum Layers: Enduring Enigma or Mystery Solved?” *Annual Review of Marine Science* 7: 207–239.
- Dang, Y.-R., Q. Q. Cha, S. S. Liu, et al. 2024. “Phytoplankton-Derived Polysaccharides and Microbial Peptidoglycans Are Key Nutrients for Deep-Sea Microbes in the Mariana Trench.” *Microbiome* 12, no. 1: 77.
- De Corte, D., S. Muck, J. Tiroch, C. Mena, G. J. Herndl, and E. Sintes. 2021. “Microbes Mediating the Sulfur Cycle in the Atlantic Ocean and Their Link to Chemolithoautotrophy.” *Environmental Microbiology* 23, no. 11: 7152–7167.
- Deutschmann, I. M., E. Delage, C. R. Giner, et al. 2024. “Disentangling Microbial Networks Across Pelagic Zones in the Tropical and Subtropical Global Ocean.” *Nature Communications* 15, no. 1: 126.
- Dlugosch, L., A. Poehlein, B. Wemheuer, et al. 2022. “Significance of Gene Variants for the Functional Biogeography of the Near-Surface Atlantic Ocean Microbiome.” *Nature Communications* 13, no. 1: 456.
- Durazzi, F., C. Sala, G. Castellani, G. Manfreda, D. Remondini, and A. de Cesare. 2021. “Comparison Between 16S rRNA and Shotgun

- Sequencing Data for the Taxonomic Characterization of the Gut Microbiota." *Scientific Reports* 11, no. 1: 3030.
- Edgar, R. C. 2010. "Search and Clustering Orders of Magnitude Faster Than BLAST." *Bioinformatics* 26, no. 19: 2460–2461.
- Edgar, R. C. 2013. "UPARSE: Highly Accurate OTU Sequences From Microbial Amplicon Reads." *Nature Methods* 10, no. 10: 996–998.
- Edgar, R. C. 2016. "SINTAX: A Simple Non-Bayesian Taxonomy Classifier for 16S and ITS Sequences." *bioRxiv*: 074161.
- Edgar, R. C. 2018. "Accuracy of Taxonomy Prediction for 16S rRNA and Fungal ITS Sequences." *PeerJ* 6: e4652.
- Giovannoni, S. J. 2017. "SAR11 Bacteria: The Most Abundant Plankton in the Oceans." *Annual Review of Marine Science* 9: 231–255.
- Halewood, E., K. Opalk, L. Custals, M. Carey, D. A. Hansell, and C. A. Carlson. 2022. "Determination of Dissolved Organic Carbon and Total Dissolved Nitrogen in Seawater Using High Temperature Combustion Analysis." *Frontiers in Marine Science* 9: 1061646.
- Hansell, D. A. 2005. "Dissolved Organic Carbon Reference Material Program." *Eos, Transactions American Geophysical Union* 86, no. 35: 318.
- Hansell, D. A., and H. W. Ducklow. 2003. "Bacterioplankton Distribution and Production in the Bathypelagic Ocean: Directly Coupled to Particulate Organic Carbon Export?" *Limnology and Oceanography* 48, no. 1: 150–156.
- Hansman, R. L., S. Griffin, J. T. Watson, et al. 2009. "The Radiocarbon Signature of Microorganisms in the Mesopelagic Ocean." *Proceedings of the National Academy of Sciences of the United States of America* 106, no. 16: 6513–6518.
- Herndl, G. J., and T. Reinthaler. 2013. "Microbial Control of the Dark End of the Biological Pump." *Nature Geoscience* 6, no. 9: 718–724.
- Hu, R., S. Liu, M. Saleem, et al. 2022. "Environmentally Induced Reconstruction of Microbial Communities Alters Particulate Carbon Flux of Deep Chlorophyll Maxima in the South China Sea." *Functional Ecology* 36, no. 10: 2493–2507.
- Hyatt, D., G. L. Chen, P. F. LoCascio, M. L. Land, F. W. Larimer, and L. J. Hauser. 2010. "Prodigal: Prokaryotic Gene Recognition and Translation Initiation Site Identification." *BMC Bioinformatics* 11: 119.
- Kanehisa, M., Y. Sato, and K. Morishima. 2016. "BlastKOALA and GhostKOALA: KEGG Tools for Functional Characterization of Genome and Metagenome Sequences." *Journal of Molecular Biology* 428, no. 4: 726–731.
- Karl, D. M., D. V. Hebel, K. Björkman, and R. M. Letelier. 1998. "The Role of Dissolved Organic Matter Release in the Productivity of the Oligotrophic North Pacific Ocean." *Limnology and Oceanography* 43, no. 6: 1270–1286.
- Khachatryan, L., R. H. de Leeuw, M. E. M. Kraakman, et al. 2020. "Taxonomic Classification and Abundance Estimation Using 16S and WGS—A Comparison Using Controlled Reference Samples." *Forensic Science International: Genetics* 46: 102257.
- King, G. M., and C. F. Weber. 2007. "Distribution, Diversity and Ecology of Aerobic CO<sub>2</sub>-Oxidizing Bacteria." *Nature Reviews. Microbiology* 5, no. 2: 107–118.
- Kolde, R. 2019. "Pheatmap: Pretty Heatmaps." R Package. Version 1.0, 8.
- Könneke, M., D. M. Schubert, P. C. Brown, et al. 2014. "Ammonia-Oxidizing Archaea Use the Most Energy-Efficient Aerobic Pathway for CO<sub>2</sub> Fixation." *Proceedings of the National Academy of Sciences of the United States of America* 111, no. 22: 8239–8244.
- Lahti, L., and S. Shetty. 2019. "Microbiome R Package." Bioconductor.
- Langmead, B., and S. L. Salzberg. 2012. "Fast Gapped-Read Alignment With Bowtie 2." *Nature Methods* 9, no. 4: 357–359.
- Li, H., B. Handsaker, A. Wysoker, et al. 2009. "The Sequence Alignment/Map Format and SAMtools." *Bioinformatics* 25, no. 16: 2078–2079.
- Liu, R., L. Wang, Q. Liu, et al. 2018. "Depth-Resolved Distribution of Particle-Attached and Free-Living Bacterial Communities in the Water Column of the New Britain Trench." *Frontiers in Microbiology* 9: 625.
- Lockwood, S., C. Greening, F. Baltar, and S. E. Morales. 2022. "Global and Seasonal Variation of Marine Phosphonate Metabolism." *ISME Journal* 16, no. 9: 2198–2212.
- Logares, R., S. Sunagawa, G. Salazar, et al. 2014. "Metagenomic 16S rDNA Illumina Tags Are a Powerful Alternative to Amplicon Sequencing to Explore Diversity and Structure of Microbial Communities." *Environmental Microbiology* 16, no. 9: 2659–2671.
- López-Pérez, M., J. M. Haro-Moreno, F. H. Coutinho, M. Martínez-García, and F. Rodríguez-Valera. 2020. "The Evolutionary Success of the Marine Bacterium SAR11 Analyzed Through a Metagenomic Perspective." *mSystems* 5, no. 5: e00605-20.
- Louca, S., L. W. Parfrey, and M. Doebeli. 2016. "Decoupling Function and Taxonomy in the Global Ocean Microbiome." *Science* 353, no. 6305: 1272–1277.
- Love, M. I., W. Huber, and S. Anders. 2014. "Moderated Estimation of Fold Change and Dispersion for RNA-Seq Data With DESeq2." *Genome Biology* 15, no. 12: 550.
- Luria, C. M., L. A. Amaral-Zettler, H. W. Ducklow, D. J. Repeta, A. L. Rhyne, and J. J. Rich. 2017. "Seasonal Shifts in Bacterial Community Responses to Phytoplankton-Derived Dissolved Organic Matter in the Western Antarctic Peninsula." *Frontiers in Microbiology* 8: 2117.
- Maidak, B. L., J. R. Cole, T. G. Lilburn, et al. 2001. "The RDP-II (Ribosomal Database Project)." *Nucleic Acids Research* 29, no. 1: 173–174.
- Marañón, E., F. van Wambeke, J. Uitz, et al. 2021. "Deep Maxima of Phytoplankton Biomass, Primary Production and Bacterial Production in the Mediterranean Sea." *Biogeosciences* 18, no. 5: 1749–1767.
- Martín-Cuadrado, A. B., P. López-García, J. C. Alba, et al. 2007. "Metagenomics of the Deep Mediterranean, a Warm Bathypelagic Habitat." *PLoS One* 2, no. 9: e914.
- McMurdie, P. J., and S. Holmes. 2013. "Phyloseq: An R Package for Reproducible Interactive Analysis and Graphics of Microbiome Census Data." *PLoS One* 8, no. 4: e61217.
- Mende, D. R., I. Letunic, J. Huerta-Cepas, et al. 2017. "proGenomes: A Resource for Consistent Functional and Taxonomic Annotations of Prokaryotic Genomes." *Nucleic Acids Research* 45, no. D1: D529–D534.
- Menzel, P., K. L. Ng, and A. Krogh. 2016. "Fast and Sensitive Taxonomic Classification for Metagenomics With Kaiju." *Nature Communications* 7, no. 1: 11257.
- Merbt, S. N., D. A. Stahl, E. O. Casamayor, E. Martí, G. W. Nicol, and J. I. Prosser. 2012. "Differential Photoinhibition of Bacterial and Archaeal Ammonia Oxidation." *FEMS Microbiology Letters* 327, no. 1: 41–46.
- Milke, F., I. Wagner-Doebler, G. Wienhausen, and M. Simon. 2022. "Selection, Drift and Community Interactions Shape Microbial Biogeographic Patterns in the Pacific Ocean." *ISME Journal* 16, no. 12: 2653–2665.
- Moran, M. A. 2015. "The Global Ocean Microbiome." *Science* 350, no. 6266: aac8455.
- Moran, M. A., E. B. Kujawinski, A. Stubbins, et al. 2016. "Deciphering Ocean Carbon in a Changing World." *Proceedings of the National Academy of Sciences of the United States of America* 113, no. 12: 3143–3151.
- Morris, R. M., M. S. Rappé, S. A. Connon, et al. 2002. "SAR11 Clade Dominates Ocean Surface Bacterioplankton Communities." *Nature* 420, no. 6917: 806–810.

- Nagata, T., H. Fukuda, R. Fukuda, and I. Koike. 2000. "Bacterioplankton Distribution and Production in Deep Pacific Waters: Large-Scale Geographic Variations and Possible Coupling With Sinking Particle Fluxes." *Limnology and Oceanography* 45, no. 2: 426–435.
- Nunoura, T., Y. Takaki, M. Hirai, et al. 2015. "Hadal Biosphere: Insight Into the Microbial Ecosystem in the Deepest Ocean on Earth." *Proceedings of the National Academy of Sciences of the United States of America* 112, no. 11: E1230–E1236.
- Nurk, S., D. Meleshko, A. Korobeynikov, and P. A. Pevzner. 2017. "metaSPAdes: A New Versatile Metagenomic Assembler." *Genome Research* 27, no. 5: 824–834.
- Oksanen, J., G. L. Simpson, F. G. Blanchet, et al. 2018. "Vegan: Community Ecology Package." <https://cran.r-project.org/web/packages/vegan/>.
- O'Leary, N. A., M. W. Wright, J. R. Brister, et al. 2015. "Reference Sequence (RefSeq) Database at NCBI: Current Status, Taxonomic Expansion, and Functional Annotation." *Nucleic Acids Research* 44, no. D1: D733–D745.
- Orellana, M. V., and D. A. Hansell. 2012. "Ribulose-1,5-Bisphosphate Carboxylase/Oxygenase (RuBisCO): A Long-Lived Protein in the Deep Ocean." *Limnology and Oceanography* 57, no. 3: 826–834.
- Ortmann, A. C., and T. T. Santos. 2016. "Spatial and Temporal Patterns in the Pelagibacteraceae Across an Estuarine Gradient." *FEMS Microbiology Ecology* 92, no. 9: fiw133.
- Reinthal, T., H. M. van Aken, and G. J. Herndl. 2010. "Major Contribution of Autotrophy to Microbial Carbon Cycling in the Deep North Atlantic's Interior." *Deep Sea Research Part II: Topical Studies in Oceanography* 57, no. 16: 1572–1580.
- Reintjes, G., C. Arnosti, B. Fuchs, and R. Amann. 2019. "Selfish, Sharing and Scavenging Bacteria in the Atlantic Ocean: A Biogeographical Study of Bacterial Substrate Utilisation." *ISME Journal* 13, no. 5: 1119–1132.
- Salazar, G., L. Paoli, A. Alberti, et al. 2019. "Gene Expression Changes and Community Turnover Differentially Shape the Global Ocean Metatranscriptome." *Cell* 179, no. 5: 1068–1083.e21.
- Salter, I., P. E. Galand, S. K. Fagervold, et al. 2015. "Seasonal Dynamics of Active SAR11 Ecotypes in the Oligotrophic Northwest Mediterranean Sea." *ISME Journal* 9, no. 2: 347–360.
- Schlitzer, R. 2023. "Ocean Data View, odv.awi.de."
- Schöne, C., A. Poehlein, N. Jehmlich, et al. 2022. "Deconstructing *Methanosarcina acetivorans* Into an Acetogenic Archaeon." *Proceedings of the National Academy of Sciences of the United States of America* 119, no. 2: e2113853119.
- Shen, Y., and R. Benner. 2018. "Mixing It Up in the Ocean Carbon Cycle and the Removal of Refractory Dissolved Organic Carbon." *Scientific Reports* 8, no. 1: 2542.
- Sintes, E., K. Bergauer, D. De Corte, T. Yokokawa, and G. J. Herndl. 2013. "Archaeal amoA Gene Diversity Points to Distinct Biogeography of Ammonia-Oxidizing Crenarchaeota in the Ocean." *Environmental Microbiology* 15, no. 5: 1647–1658.
- Srivastava, A., D. de Corte, J. A. L. Garcia, et al. 2023. "Interplay Between Autotrophic and Heterotrophic Prokaryotic Metabolism in the Bathypelagic Realm Revealed by Metatranscriptomic Analyses." *Microbiome* 11, no. 1: 239.
- Sunagawa, S., L. P. Coelho, S. Chaffron, et al. 2015. "Ocean Plankton. Structure and Function of the Global Ocean Microbiome." *Science* 348, no. 6237: 1261359.
- Swan, B. K., M. Martinez-Garcia, C. M. Preston, et al. 2011. "Potential for Chemolithoautotrophy Among Ubiquitous Bacteria Lineages in the Dark Ocean." *Science* 333, no. 6047: 1296–1300.
- Thrash, J. C., B. Temperton, B. K. Swan, et al. 2014. "Single-Cell Enabled Comparative Genomics of a Deep Ocean SAR11 Bathytype." *ISME Journal* 8, no. 7: 1440–1451.
- UNESCO. 1994. *Protocols for the Joint Global Ocean Flux Study (JGOFS) Core Measurements*, 170. UNESCO-IOC.
- Vergin, K. L., B. Beszteri, A. Monier, et al. 2013. "High-Resolution SAR11 Ecotype Dynamics at the Bermuda Atlantic Time-Series Study Site by Phylogenetic Placement of Pyrosequences." *ISME Journal* 7, no. 7: 1322–1332.
- Vihtakari, M. 2024. "ggOceanMaps: Plot Data on Oceanographic Maps Using 'ggplot2'." R Package Version 2.2.0.
- Wright, C. L., and L. E. Lehtovirta-Morley. 2023. "Nitrification and Beyond: Metabolic Versatility of Ammonia Oxidising Archaea." *ISME Journal* 17, no. 9: 1358–1368.
- Xue, C. X., J. Liu, D. J. Lea-Smith, et al. 2020. "Insights Into the Vertical Stratification of Microbial Ecological Roles Across the Deepest Seawater Column on Earth." *Microorganisms* 8, no. 9: 1309.

### Supporting Information

Additional supporting information can be found online in the Supporting Information section. **Figure S1:** Venn diagram of the prokaryotic community's core members across different depth layers (a). Taxonomic affiliation of the prokaryotic core members (b). **Figure S2:** Heatmap of the detected genes (KOs) across the longitudinal transect and the entire water column. **Figure S3:** Spearman correlation between indexes (diversity, richness and evenness indexes from mOTUS and KOs) and different physical-chemical and biological variables. **Figure S4:** DOC, mOTUS and KOs diversity plots throughout the longitudinal transect in the whole depth profile.